

The Effects of Phytobiotic-Enriched Diet on Immunity Index and Hematological-Biochemical Changes in Common Carp Fish (*Cyprinus carpio*)

Seyede Rahime Razavi Amria, Hosna Gholipour Kanania, Mohammad Farhangi* and Hosein Adineha

^aDepartment of Fisheries, Faculty of Agriculture and Natural Resources, Gonbad Kavous University, Gonbad Kavous, Golestan, Iran

*Corresponding author email: farhangi@gonbad.ac.ir

Article History: Received: 29 April 2023/Accepted in revised form: 21 July 2023

© 2012 Iranian Society of Medicinal Plants. All rights reserved

ABSTRACT

The present study compared the effects of four medicinal herbs on the growth, digestive enzyme activity, innate immunity, and oxidative status of common carp juveniles ($Cyprinus\ carpio$). The juvenile fish (average weight: $80.01\pm0.34\ g$) were fed with four diets, including $1\%\ Zataria\ multiflora$ oil (T2), Zingiber powder (T3), garlic essential oil extract (T4), garlic peel extract (T5), and control diet (T1) without any additives, for seven weeks (20 individuals per replicate). The results showed that weight gain and specific growth rate were highest in the treatment fed with garlic and ginger extracts (T3 and T4). The protease and lipase activities in the medicinal herb treatments (especially the garlic extract treatment; T4) were significantly higher than in the control. Fish fed with medicinal herb-supplemented diets (except for $Zataria\ multiflora\ oil$; T2) also displayed higher levels of serum lysozyme, alternative complement, total immunoglobulin, superoxide dismutase, and total protein levels compared to the control diet. The highest levels were observed at the 1% level of Zingiber powder (T3) and garlic extract (T4). Medicinal herb supplementation at a level of 1% is a natural immunostimulant and growth promoter supplement recommended for common carp.

Keyword: Cyprinus carpio, Medicinal plants, Nutrition, Health.

INTRODUCTION

Common carp (*Cyprinus carpio*) is an important species for aquaculture in many Asian and some European countries. It is commonly found in freshwater environments such as rivers, ponds, and lakes, and is seldom found in brackish water environments [1]. The use of antimicrobials and sanitizers has shown partial success in preventing or treating marine ailments. Currently, herbal and natural remedies have a particular position in the treatment of ailments due to their commercial value, low production cost, ecological friendliness (organic drugs), minimal side effects compared to chemical medicines, absence of pathogen resistance, the uniqueness of disease treatment with plants, and various medicinal skills regarding pharmaceutical plants [2]. Therefore, exploring different food additives is a very important goal for aquaculture researchers [3]. Garlic (*Allium sativum* L.) is a medicinal plant that has been used in Indian Ayurvedic medicine for a thousand years. Garlic has shown antimicrobial, antihypertensive, hepatoprotective, antioxidant, and immune

enhancing properties, including the promotion of lymphocyte proliferation, cytokine release, phagocytosis, and natural killer cell activity [4]. Ginger (*Zingiber officinale*) is another potential medicinal plant that contains natural antioxidants such as gingerols, shogaols, and zingerones that can improve disease resistance against pathogens [5]. *Zataria multiflora*, from the family Labiatae, is traditionally used as an antiseptic, anesthetic, and antispasmodic [6]. In Iran, it is more widely used as an herbal tea, flavoring agent (condiment and spice), and medicinal plant. It is also used as a tonic, carminative, digestive, antispasmodic, anti-inflammatory, antitussive, expectorant, and for the treatment of colds in Iranian traditional medicine [7]. Improving the immune system of fish is considered the most effective method of preventing fish diseases in aquaculture. This improvement can be achieved through the use of vaccines, which enhance the specific immune response of fish and are considered the most effective agents. As the concern about antibiotic resistance in aquaculture grows, there has been a rising trend towards the use of ancient herbal medications. The tendency towards ancient herbal medications is on the

rise in aquaculture due to concerns about antibiotic resistance. According to several applied research studies in this area, herbal medicine has shown a potential capacity to boost the immune function of aquatic animals contributing to disease resistance enhancement [8]. Several applied research studies have shown that herbal medicine has the potential to boost the immune function of aquatic animals, which can contribute to enhancing disease resistance [9, 10, 11, 12, 13, 14 and 15]. Little information is available on the effects of garlic peel in aquaculture [13]. Therefore, this study aims to compare the effects of various herbal medicines on the growth performance, hematological factors, and intestinal enzymes of *C. carpio*.

MATERIAL AND METHODS

Animals

A total of 300 common carp specimens (*Cyprinus carpio*) were obtained from a carp farm located in Mazandaran province, Iran. The fish were transported to the Aquatic Animal Health Laboratory at Gonbad Kavous University, Iran. The fish were then acclimated to laboratory conditions for 10 days in fiberglass-reinforced plastic tanks and fed a commercial diet. The laboratory conditions were regularly monitored and maintained at a consistent level during the acclimation period.

Treatments

For this study, five treatments (T1, T2, T3, T4, and T5) with three replicates were established. The control group was fed a basic diet (T1). *Zataria multiflora* oil (T2) and *Zingiber officinale* powder (T3) were added to the other experimental diets. The remaining groups were fed garlic extract (T4) and garlic peel extract (T5). The ingredients were added at a concentration of 1% [16, 17] to each experimental diet using 5% gelatin in sufficient water as a binder [18]. Each replicate consisted of 20 fish $(80.01 \pm 0.34 \text{ g})$ in a 200-liter tank with 120 liters of water at a salinity level of 0.56 ± 0.03 ppt [19]. Aeration was provided by a single air stone to maintain the dissolved oxygen level at 7.72 ± 0.18 mg/l. The water temperature was maintained at 25.19 ± 0.08 °C and pH 7.08 ± 0.05 throughout the experimental period. The control diet was provided by enhancing the water with no crust power. The fish were fed 2.5% of their average body weight per day for seven weeks, divided into three feedings at 08:00, 11:00, and 18:00. Feces, molts, and dead fish were removed daily, and 30% of the water was changed with fresh water in each tank [5].

Plants Management

Garlic was purchased from a local marketplace. The garlic cloves and peels were dried at 37°C for three days and ground into a fine powder. The dried powders were soaked in a mixture of water and ethanol (1:1 ratio) for 48 hours. The extraction was filtered (Whatman No.1) and concentrated at 40°C using a rotary evaporator (HS-200S, Korea). Finally, the medicinal plant extracts were stored at 4°C until use [20].

Ginger (*Zingiber officinale*) was also purchased from a local marketplace and powdered for use in the diet. *Zataria multiflora* essential oil was prepared by Barij-Essence Company, Iran. The diets were then dried, sealed in plastic bags, and stored at 4°C until feeding [3, 21]. The proximate composition of the experimental diets (crude protein, crude lipid, crude ash, and crude fiber) was determined using AOAC [22] (Table 1).

Growth and Nutrition Parameters

At the end of the study, the final body weight (FBW), final body length (FBL), condition factor (CF), protein efficiency ratio (PER), lipid efficiency ratio (LER), feed conversion efficiency (FCE), specific growth rate (SGR), feed conversion ratio (FCR), and survival rate were calculated for both the experimental and control groups. The following formulas were used to calculate the growth factors [4]:

Final body weight (FBW) = Mean weight of fish at the end of the study Final body length (FBL) = Mean length of fish at the end of the study Condition factor (CF) = (Fish weight (g) / Fish length (cm)3) \times 100 Protein efficiency ratio (PER) = (Weight gain (g) / Protein intake (g)) Lipid efficiency ratio (LER) = (Weight gain (g) / Lipid intake (g)) Feed conversion efficiency (FCE) = (Weight gain (g) / Feed intake (g)) \times 100 Specific growth rate (SGR) = $[(ln(final\ weight) - ln(initial\ weight)) / Time\ (days)] \times 100$ Feed conversion ratio (FCR) = Feed intake (g) / Weight gain (g) Survival rate = (Number of surviving fish / Initial number of fish) × 100

Table 1 Ingredients and Composition of the Experimental Diets

Ingredient	(g kg ⁻¹)	Composition	%
Fishmeal ¹	150	Dry matter	85.2
Poultry meal ²	200	Crude protein	37.2
Soybean meal	200	Crude lipid	7.8
Wheat flour	378	Ash	6.2
Fish oil	15		
Soybean oil	15		
Corn flour	30		
L-Lysine ³	4		^
L-Methionine	6		
Vitamin premix a	1		A', C)
Mineral premix b	1		

- 1- Pars kilka Co., Mazandaran, Iran (Kilka powder analysis; Protein: 70–72%, Fat: 8–11%, Ash: 11.6%, Moisture: 7–9%).
- 2- Makianmehr Co., Golestan, Iran. 3- Morghenojan.Co., Tehran, Iran.
- a Vitamin premix (per kg of diet): vitamin A, 2000 IU; vitamin B1 (thiamin), 5 mg; vitamin B2 (riboflavin), 5 mg; vitamin B6, 5 mg; vitamin B12, 0.025 mg; vitamin D3, 1200 IU; vitamin E, 63 mg; vitamin K3, 2.5 mg; folic acid, 1.3 mg; biotin, 0.05 mg; pantothenic acid calcium, 20 mg; inositol, 60 mg; ascorbic acid (35%), 110 mg; niacinamide, 25 mg.
- b Mineral premix (per kg of diet): MnSO4, 10 mg; MgSO4, 10 mg; KCl, 95 mg; NaCl, 165 mg; ZnSO4, 20 mg; KI, 1 mg; CuSO4, 12.5 mg; FeSO4, 105 mg; Co, 1.5 mg.

Sampling and Processing

After seven weeks of feeding, the fish were fasted for 24 hours before blood and serum collection. Nine fish from each treatment group were randomly selected for blood sampling from the caudal veins [4]. The sampled blood was then centrifuged at 2000 g (at 4°C) for 10 minutes. Additionally, 12 fish were randomly selected, euthanized, and dissected to collect the entire digestive tract [23]. The serum and intestine were extracted and frozen at -80°C for serum and digestive enzyme analyses [24].

Digestive Enzyme Parameters

The digestive tract samples were homogenized, centrifuged at 25000 g for 20 minutes, and the supernatants were collected [25]. Amylase activity was measured using the method described by Langlois et al. [26] with 0.3% soluble starch as the substrate dissolved in NaH2PO4 buffer (pH 7.4). Protease activity was determined following the method described by Walter [27], using casein (Sigma) at a concentration of 1% w/v as the substrate in 0.2 M phosphate buffer at pH 7.0. To measure the lipase activity, p-nitrophenol myristate was used as the substrate dissolved in 0.25 M Tris-HCl (pH 9.0) [28]. These parameters were used to assess the effects of the different herbal additives on the digestive enzyme activity of *C. carpio*.

Immune and Biochemical Parameters

The lysozyme levels were determined using the method described by Ellis [29]. The total immunoglobulin was evaluated according to the Siwicki and Anderson method [30]. Alternative hemolytic complement activity (ACH50) was defined based on the hemolysis of rabbit red blood cells (RaRBC) using the Sunyer and Tort method [31]. The content of alternative complement activity (ACH50) was measured and used to calculate the complement activity of the samples (the value of ACH50 is in units per ml). Serum total protein (TP) and glucose were measured using Parsazmon's kits (Parsazmon Company, Iran) based on the company's protocol.

Antioxidant Responses

Superoxide dismutase (SOD) activity was determined using the method described by Fridovich [32] which is based on the inhibition of nitroblue tetrazolium reduction by the xanthine / xanthine oxidase system as a

superoxide generator. Catalase (CAT) activity was measured according to the method described by Beutler [33]. Malondialdehyde (MDA) levels, which are an indicator of free radical generation, were estimated according to Lowry et al. [34].

Data Analyses

The normality of the data was determined using the Shapiro-Wilk Test. Significant differences between treatments were evaluated using One-way analysis of variance (ANOVA). Duncan's test was used at a significant level of (P< 0.05) to compare means (Mean \pm SEM). Statistical analyses were performed using SPSS 21 and Excel 2013 software.

RESULTS

Growth Performance

The effect of the experimental diets on the growth performance of common carps ($C.\ carpio$) is shown in Table 2. The final weight of the fish showed significant differences among the groups (P < 0.05). The highest significant final weights were recorded in the ginger and garlic groups compared to the control and other experimental groups. The specific growth rate (SGR) was significantly increased (P < 0.05) in all groups compared to the control group. Fish fed with ginger and garlic showed the highest SGR compared to other experimental groups. All the fish fed with the experimental diets had lower feed conversion ratio (FCR) compared to the control group. The lowest FCR was recorded in the fish fed with the ginger and garlic diet compared to other experimental groups. Additionally, the fish receiving diets supplemented with ginger and garlic had significantly higher growth parameters, including condition factor (CF) as well as protein efficiency ratio (PER) (P < 0.05). The survival rate was not affected (P > 0.05) by the herbal-supplemented diets. These results suggest that the addition of ginger and garlic to the diets of $C.\ carpio$ can improve their growth performance. (Table 2).

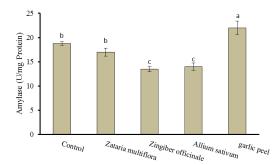
Table 2 Growth and Feed Performance of Common Carp Fed with Medicinal Plants after 7 Weeks (Mean ± SEM)

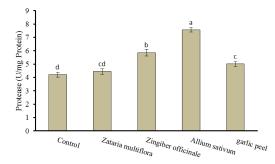
	Treatments				
Growth	T1	T2	T3	T4	T5
parameters	(Control)	(garlic extract)	(garlic peel)	(Z. multifora oil)	(Z. officinale powder)
FBW (gr)	115.09±0.47 c	129.18±0.67 a	121.95±0.97 b	120.29±0.38 b	131.81±0.57 a
CF (gr/cm ³)	0.91±0.04 ab	1.24±0.07 a	1.13±0.05 ab	1.18±0.06 a	1.37±0.05 b
SGR (%/day)	0.78±0.02 c	0.94±0.017 a	0.85±0.026 b	0.86±0.019 b	0.94±0.017 a
FCE (gr/cm ³)%	23.65±9.50 d	65.39±9.22 b	43.96±13.25 c	39.03±5.24 c	73.19±7.78 a
FCR	2.75±0.06 a	2.11±0.03 c	2.53±0.08 b	2.48±0.06 b	2.09±0.03 c
PER (gr/gr)	1.05±0.024 c	1.35±0.023 b	1.15±0.03 4c	1.16±0.025 c	1.47±0.02 6a
LER (gr/gr)	1.33±0.13 c	3.68±0.14 a	2.49±0.20 b	2.15±0.13 b	4.13±0.13 a
Survival (%)	100	100	100	100	100

^{*} Final body weight (FBW), Condition factor (CF), Specific growth rate (SGR), Feed conversion efficiency (FCE), Feed conversion ratio (FCR), Protein efficiency ratio (PER), Lipid efficiency ratio (LER)

Digestive Enzymes

The activities of digestive enzymes in common carp fed diets supplemented with 1% of medicinal plants for 7 weeks are shown in figure 1. The activity of amylase was significantly affected by the ginger and garlic groups. The protease and lipase activities were significantly higher in fish fed *Allium sativum* (garlic extract) and *Zingiber officinale* (ginger), respectively (figure 1).





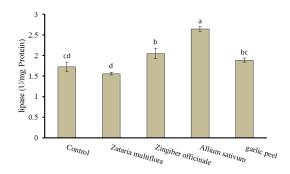


Fig. 1 Digestive Enzyme Activities of Juvenile Common Carp Fed Medicinal Plants for 7 Weeks (Mean \pm SEM). Different Lowercase Letters within a Column Show Significant Effects of the Treatments (P < 0.05).

Immune and Biochemical Response

After 7 weeks, the serum lysozyme activity and Total immunoglobulin of common carps in the ginger, garlic, and garlic peel treatments were significantly higher (p < 0.05) than those in the control group (Fig 2). The treatments inoculated with garlic and ginger exhibited the highest total immunoglobulin level (P < 0.05). However, the Total immunoglobulin was not statistically significant (P > 0.05) between the ginger and garlic groups (P < 0.05). The carps' alternative hemolytic complement activity (ACH50) was influenced (P < 0.05) by ginger, garlic, and garlic peels as food additives compared to the control group. However, a numerically higher ACH50 level was found in the fish receiving ginger. The glucose level was significantly increased in those treated with garlic peel (P < 0.05). The glucose level of fish after 4 weeks of experimental diet was significantly increased in fish treated with garlic peel (P < 0.05) (Fig 2).

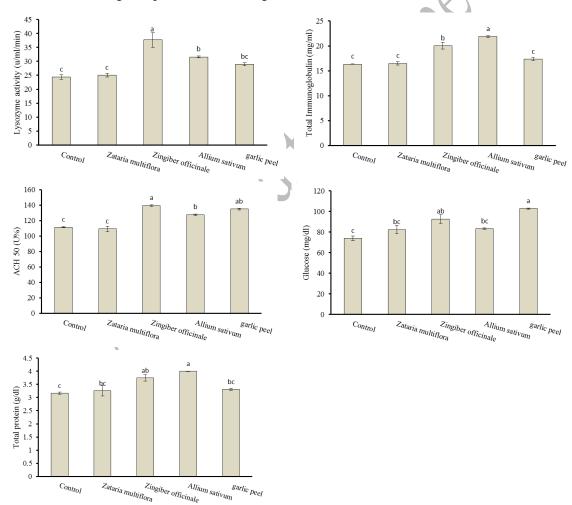


Fig. 2 Innate Immune Responses of Common Carp Fed Medicinal Plants for 7 Weeks (Mean \pm SEM). Different Lowercase Letters within a Column Show Significant Effects of the Treatments (P < 0.05).

Antioxidant Enzymes Activities

After 7 weeks of oral administration of four different herbal diets, the serum SOD activity of common carps in the ginger and garlic treatments was significantly higher (p < 0.05) than in the *Zataria multiflora* and control groups (Fig 3). Catalase activity of fish fed with ginger and garlic treatment diets was significantly (P < 0.05) higher than the other treatments. The MDA level was decreased significantly in fish fed medicinal plants compared to the control group, with the lowest being in fish fed with ginger (Z. officinale) diet (Fig 3).

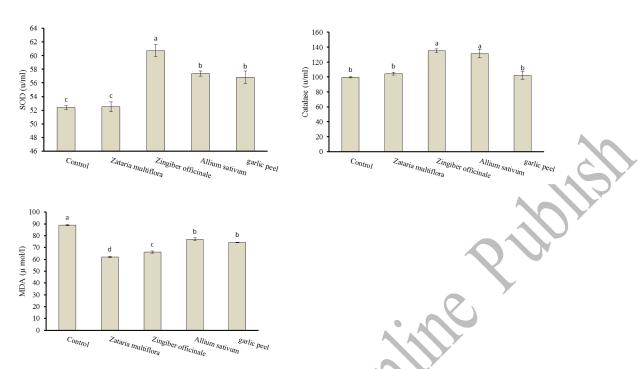


Fig. 3 Antioxidant Enzyme Activities of Common Carp Fed Medicinal Plants for 7 Weeks (Mean \pm SEM). Different Lowercase Letters within a Column Show Significant Effects of the Treatments (P < 0.05).

DISCUSSION

Hematological indicators are an important criterion for evaluating the effects of dietary treatments on animals [13]. For instance, Thanikachalam et al. [35] reported a significant increase in the WBC count of African catfish (Clarias gariepinus) following a 20-day post-feeding with garlic peel. Chitsaz et al. [11] also reported significant increases in the Hb level in H. huso fed with a diet incorporated with garlic peel compared to the control treatment. The use of immunostimulants in fish diets as potential protective food additives has been reviewed by Wang et al. [36]. In the present study, the introduction of herbal additives to the fish diet improved growth performance, including WG, SGR, FCR, PER, and LER, notably in the ginger and garlic treatments, followed by garlic peel and Zataria multiflora, compared to the control group. Consistent with our study, garlic powder supplementation administered dietary to common carps could enhance growth parameters. Norhan et al. [37] demonstrated that 1% garlic administration improved growth rate and feed utilization in sea bass. Conversely, no significant variations were detected in the body weight of tilapia fed with dietary supplements of 0.5% and 1% garlic for four weeks [38]. Digestion is an important process in animal metabolic rate because it regulates the availability of nutrients required for all biotic purposes and is the main instrument in considering the dietary situation and conformity of the creature to nutritional variation [11]. It has been revealed that medicinal plants stimulate the secretion of pancreatic enzymes, which are imperative parameters in nutrient breakdown and absorption [39]. According to this study, carps fed with garlic and garlic peel diets showed enhanced activity of digestive enzymes such as protease and lipase, which improved digestion and intake of essential nutrients for fish development. These results are similar to the findings of Chitsaz et al. [11], who reported increased digestive enzyme activity in Huso huso fed with garlic peel. Similarly, many researchers have reported improved amylase activity under various herbal additives [40]. Jang et al. [41] also demonstrated that herbal medicine administration could enhance the pancreatic digestive enzyme activities, such as α-amylase and trypsin, as well as intestinal enzymes, improving the capability to enhance fish growth efficiency [42]. Moreover, this improvement is potentially due to the development of intestinal microflora, contributing to a decrease in pathogenic bacteria growth and an increase in the colonization of beneficial bacteria [43, 44]. Therefore, the use of medicinal plants can improve the digestive process, leading to better nutrient absorption and utilization, ultimately improving the growth performance of aquatic animals.

Humoral and cellular immunity are the specific defense mechanisms in fish. Cellular immunity includes neutrophils, phagocytic cells, natural killer cells, and lymphocytes, while humoral immunity includes immunoglobulins, hemolysins, lysozyme, and complement molecules [42]. In the present study, garlic and ginger treatments positively affected the total protein concentration in common carps. Similarly, garlic and ginger oil increased the total protein concentration in sea bass [45].

Moreover, the supplementation of fish diets with ginger extracts has been shown to increase TP levels in fish plasma [46]. Similarly, powdered or oil forms of garlic have been found to improve TP in the blood of Oreochromis niloticus [47, 48]. An increase was observed in the alternative complement activity (ACH50) in carps fed with ginger, garlic, and garlic peel, potentially improving phagocytosis by enhancing the recognition and elimination of bacterial agents. Several studies have reported enhancement of complement activity following the administration of different immunostimulants such as herbal derivatives [49]. Serum immunoglobulin mainly neutralizes exogenous pathogens and is considered a component of the teleost humoral immune system. In this study, feeding fish with ginger and garlic-supplemented diets resulted in higher levels of immunoglobulin in carps compared to the control. Sarhadi et al. [50] also reported that artemisia (Artemisia annua) leaf extract improved the immune response, including lysozyme, immunoglobulin, and protease. The remarkable enhancement in serum immunoglobulin levels may be due to the active ingredients in garlic (known as allicin), which can actively stimulate the secretion of IgM. Lysozymeis the first line of defense in the innate immune system [51]. Its function is to act as a marker of innate immunity in leukocyte respiratory burst activity [52]. Blood lysozyme actively lyses peptidoglycans in the bacterial cell wall, and it is considered an important factor in the defense against bacterial infections. In the present study, garlic (1%) significantly enhanced lysozyme activity, suggesting that the use of medicinal plants can improve the innate immune response of aquatic animals. Additionally, lysozyme works as an opsonin and activates phagocytes and the complement system, further enhancing the immune response [3].

Free radicals are produced during the oxidative stress process to protect living organisms from invading pathogens. However, these free radicals can be harmful to living organisms themselves, and an antioxidant system is applied to protect them from free radicals. The most important indicators of antioxidant activities are the increase in the SOD and CAT enzyme levels [53, 54, 55]. SOD is the first line of defense against oxygen toxicity, using its inhibitory effects on oxyradical formation and contributing to the dismutation of the superoxide anion radical into water and hydrogen peroxide, which is then detoxified by CAT [56]. In the present study, fish treated with ginger and garlic diet had the highest SOD and catalase levels. In two similar studies on tilapia, garlic and ginger were found to increase antioxidant enzyme activity (SOD and CAT) [57, 58, 59]. In other research, the addition of purslane extract (*Portulaca oleracea*) to the diet of grass carp (*Ctenopharyngodon idella*) led to a statistically significant increase compared to the control [60]. In general, the results show that ginger reduced the oxidative stress caused by stress by inhibiting radicals, because the highest level of superoxide dismutase (SOD) and the lowest level of malondialdehyde (MDA) were observed in the treatment fed with ginger [16].

Malondialdehyde (MDA) is one of the end products of lipid peroxidation caused by oxygen free radicals, and it is an important oxidative stress indicator that shows the degree of lipid peroxidation. An increase in MDA levels is an important indicator of cellular damage [61]. In this study, low MDA values were obtained in all experimental groups, especially in ginger-treated fish, when compared to the control. Similarly, it has been reported by Sahan et al. [59] and Islam et al. [58] that MDA levels decreased in tilapia fed ginger. Our results were also in accordance with the findings of Giannenas et al. [62] in the ginger-fed group. It has been emphasized that this maybe associated with various bioactive substances (shagol, gingerols, zingeron, etc.) found in the composition of ginger, which provide antioxidant properties to ginger. Phenolic compounds of ginger (gingerols, shogaols, volatile oils, flavonoids, and phenolic ketone derivatives) have been reported to promote antioxidant activity against free radicals and inhibit lipid peroxidation [63].

CONCLUSION

Based on the results obtained in the present study for common carps fed with ginger and garlic-added diets at a 1% concentration, it can be concluded that these medicinal plants can enhance the digestive activity, growth function, and general health status of aquatic animals. The findings suggest that the use of phytobiotics, especially garlic administration in fish diets, can improve growth functions and reduce oxidative stress. These results are consistent with previous research indicating that the use of medicinal plants in aquaculture can enhance the immune response and antioxidant defense system of aquatic animals, ultimately leading to better disease resistance and improved overall health. Therefore, the use of medicinal plants such as ginger and garlic in aquaculture can be a promising and sustainable approach to improve the performance and health of aquatic animals, while also reducing the use of synthetic chemicals in aquaculture.

Ethical Statement

This work was conducted in accordance with the guidelines and regulations approved by the Ethics Committee of Gonbad Kavous University, Iran. All procedures involving animal handling and experimentation were carried out in strict accordance with the guidelines and regulations for animal welfare and protection. Special attention was given to minimize the stress and discomfort of the experimental animals throughout the study. No animals were harmed or sacrificed unnecessarily during the course of this research.

ACKNOWLEDGEMENTS

We would like to express our gratitude to Gonbad Kavous University, Golestan, Iran, for their continuous support in providing the research project.

Conflict of Interest

The authors declare that there is no potential conflict of interest related to this research.

Authors Contributions

All authors have contributed equally to this research project. They provided critical feedback, designed the study, analyzed the data, and helped shape the manuscript.

REFERENCES

- 1. Rahman M.M. Role of common carp (*Cyprinus carpio*) in aquaculture production system. Frontiers in Life Science. 2015; 8: 399-410.
- 2. Ghasemi Pirbalouti A. Iranian medicinal and aromatic plants (2nd edition). Islamic Azad University Publishers, Shahrekord, Iran. (In Persian). 238p; 2009.
- 3. Citarasu T., Sivaram V., Immanuel G., Rout N., Murugan V. Influence of selected Indian immunostimulants herbs against white spot syndrome virus (WSSV) infection in black tiger shrimp, *Penaeus monodon* with reference to Haematological, biochemical and immunological changes. Fish and Shellfish Immunology. 2006; 21: 372–384.
- 4. Adineh H., Harsij M., Jafaryan H., Asadi M. The effects of microencapsulated garlic (*Allium sativum*) extract on growth performance, body composition, immune response and antioxidant status of rainbow trout (*Oncorhynchus mykiss*) juveniles. Journal of Applied Animal Research. 2020; 48: 372-378.
- 5. Jabran A.G., Soeharto S., Sujuti H. The effect of ginger (*Zingiber officinale Roscoe*) extract on liver histopathology and alanine aminotransferase serum level in carbofuran-induced rats. International J Pharm Tech Research. 2015; 8: 889-897.
- 6. Ramezani M., Hosseinzadeh H., Samizadeh S. Antnociceptive effects of *Zataria multiflora* Boiss fractions in mice. J Ethnopharmacology. 2015; 91: 167-170.
- 7. Sepavand A., Ashraf Jafari A., Sefidkon F., Kalatejari S. Evaluation of growth, essential oil content and composition in four thyme species under dryland farming system in Zagheh, Rangeland, Khoramabad, Iran. J Medicinal Plants and Byproducts. 2021; 1: 59-65.
- 8. Buchmann K. Evolution of innate immunity: clues from invertebrates via fish to mammals. Frontiers in Immunology. 2014; 5: 459
- 9. Dadgar SH, Seidgar M., Nekuiefard A., Valipour A.R., Sharifian M., Hafezieh M. Oral administration of garlic powder (*Allium sativum*) on growth performance and survival rate of Carassius auratus fingerlings. Iranian J Fisheries Sci. 2019; 18: 71-82

- 10. Labrador J.R.P., Guinares R.C., Hontiveros GJS. Effect of garlic powder supplemented diets on the growth and survival of Pacific white leg shrimp (*Litopenaeus vannamei*). Cogent Food Agriculture. 2016; 2: 295-299.
- 11. Lokesh B., Neeraja T., Haribabu P., Ramalingaiah D., Pamanna D. Effect of garlic supplemented diets on the growth and survival of Pacific white leg shrimp (*Litopenaeus vannamei*) Juveniles. J Entomology and Zoology Studies. 2020; 8: 295-299
- 12. Tanekhy M., Fall J. Expression of innate immunity genes in kuruma shrimp *Marsupenaeus japonicus* was studied after in vivo stimulation with garlic extract (allicin). Veterinarni Medicina. 2015; 60: 39–47.
- 13. Chitsaz H., Oraji H., Keramat Amirkolaie H.A., Akrami A. Effect of garlic peel on haematological, biochemical and digestive enzyme activity in beluga juvenile (*Huso huso*). Iranian J Aquatic Animal Health. 2018; 4: 13-28.
- 14. Kesbic O.S, Acar U., Hassaan M.S, Yılmaz, S, Guerrera, M.C, Fazio F. Effects of Tomato Paste by-product extract on growth performance and blood parameters in common carp (*Cyprinus carpio*). Animal. 2022; 12: 3387.
- 15. Ngugi C.C., Oyoo-Okoth E., Mugo-Bundi J., Orina P.S., Chemoiwa E.J., Aloo P.A. Effects of dietary administration of stinging nettle (*Urtica dioica*) on the growth performance, biochemical, hematological and immunological parameters in juvenile and adult Victoria Labeo (*Labeo victorianus*) challenged with *Aeromonas hydrophila*. Fish and Shellfish Immunology. 2015; 44: 533–541.
- 16. Fazelan Z., Vatnikov Y.A., Kulikov E.V., Plushikov V.G., Yousefi M. Effects of dietary ginger (*Zingiber officinale*) administration on growth performance and stress, immunological, and antioxidant responses of common carp (*Cyprinus carpio*) reared under high stocking density. Aquaculture. 2020; 518: 734833.
- 17. Yousefi M., Zahedi S., Reverter M., Adineh H., Hoseini S.M., Van Doan H., Hoseinifar S.H. Enhanced growth performance, oxidative capacity and immune responses of common carp, *Cyprinus carpio* fed with *Artemisia absinthium* extract-supplemented diet. Aquaculture. 2020; 545: 737167.
- 18. Lee D.H., Lim S.R., Han J.J., Lee S.W., Ra C.S., Kim J.D. Effect of dietary Garlic powder on growth, feed utilization and whole body composition changes in fingerling starlet sturgeon, Acipenser ruthenus. Asian-Australasian Journal of Animal Sciences. 2014; 27: 1303-1310.
- 19. Adineh H., Naderi M., Hamidi M.K., Harsij M. Biofloc technology improves growth, innate immune responses, oxidative status, and resistance to acute stress in common carp (*Cyprinus carpio*) under high stocking density. Fish and shellfish immunology. 2019; 95: 440-448.
- 20. Arabshahi-Delouee M., Asna Urooj S. Antioxidant properties of various solvent extracts of mulberry (*Morus indica* L.) leaves Author links open overlay panel. Food Chemistry. 2007; 102: 1233-1240.
- 21. Eloff J.N. Which extractant should be used for the screening and isolation of antimicrobial components from plants? *Journal* of Ethno pharmacology. 1998; 6: 1-8.
- 22. AOAC. Official Methods of Analysis, Association of Official Analytical Chemists International, Gaithersburg, 2005.
- 23. Chang Y. P., Liu C.H., Wu C.C., Chiang C.M., Lian J.L., Hsieh S.L. Fish Dietary administration of zingerone to enhance growth, non-specific immune response, and resistance to Vibrio alginolyticus in pacific white shrimp (*Litopenaeus vannamei*) juvenles. Fish and Shell fish Immunology. 2012; 32: 284-290.
- 24. Ahmadifard N., Abedian Kenari A., Motamedzadegan A. Study of Proteases (Gastric, Intestine and Pancreas) Enzyme Activities of Rainbow Trout, *Oncorhynchus mykiss*, fed partial substitution of dietary fish meal with rice bran protein concentrate. Journal of Fisheries-Iranian. 2012; 65: 365-376.
- 25. Najdegerami E.H., Bakhshi F., Lakani F.B. Effects of biofloc on growth performance, digestive enzyme activities and liver histology of common carp (*Cyprinus carpio* L.) fingerlings in zero-water exchange system. Fish physiology and biochemistry. 2016; 42: 457-465.
- 26. Langlois A., Corring T., Fevrier C. Effects of wheat bran on exocrine pancreas secretion in the pig. Reproduction Nutrition Developpement. 1987; 27: 929-939.
- 27. Walter H. Proteinase's methods with hemoglobin, casein and azocoll as substrates. In: H. U. Bergmeyer (ed.), Methods of enzymatic analysis, 1984.
- 28. Iijima N., Tanaka S., Ota Y. Purification and characterization of bile salt-activated lipase from the hepatopancreas of red sea bream, *Pagrus major*. Fish Physiology and Biochemistry. 1998; 18: 59-69.
- 29. Ellis A. Lysozyme assay in techniques in fish immunology, SOS Publications, Fair Haven, NJ, USA, 1990.
- 30. Siwicki A.K., Anderson D.P. Nonspecific defense mechanisms assay in fish. II. Potential killing activity of neutrophils and macrophages, lysozyme activity in serum and organs and total immunoglobulin (Ig) level in serum. In: Fish diseases diagnosis and preventions methods, Siwicki A.K., Anderson D.P., Waluga J. (eds). Wydawnictwo Instytutu Rybactwa Śródladowego. 1993: 105-111.
- 31. Sunyer A., Tort E. Natural hemolytic and bactericidal activities of sea bream *Sparus aurata* serum are effected by the alternative complement pathway. Veterinary Immunology and Immunopathology. 1995; 45: 333-345.
- 32. Fridovich I. Superoxide radical and superoxide dismutases. Annual Review of Biochemistry. 1995; 64; 97-112.
- 33. Beutler E. Red cell metabolism, a manual of biochemical methods. Grune and Straton Press, New York, 1975.

- 34. Lowry O.H., Rosenbrough W.J., Fair H.L., Randall R.J. Protein measurement with folin phenol reagent. Journal of Biological Chemistry. 1951; 19: 265-275.
- 35. Thanikachalam K., Kasi M., Rathinam X. Effect of garlic peel on growth, hematological parameters and disease resistance against *Aeromonas hydrophila* in African catfish *Clarias gariepinus* (Bloch) fingerlings. Asian Pacific Journal of Tropical Medicine. 2016; 3: 614-618.
- 36. Wang A., Meng X., Lub R., Wu C., Luo Y.T., Yan X., Li X.J., Kong X.A., Nie G.X. Effects of *Rehmannia glutinosa* on growth performance, immunological parameters and disease resistance to *Aeromonas hydrophila* in common carp (*Cyprinus carpio* L.). Aquaculture. 2015; 435: 293-300.
- 37. Norhan E., SalehFady R., Michael M., Toutou R. Evaluation of garlic and onion powder as phyto-additives in the diet of sea bass (*Dicentrarcus labrax*). The Egyptian Journal Aquatic Research. 2015; 41: 211-217.
- 38. Ndong J.D., Fall J. The effect of garlic (*Allium sativum*) on growth and immune responses of hybrid tilapia (*Oreochromis niloticus*×*Oreochromis aureus*). Journal of Clinical Immnunol and Immunopathology Research. 2011; 3: 1-9.
- 39. Frankic T., Voljc M., Salobir Rezar V. Use of herbs and spices and their extracts in animal nutrition. Acta agriculturae Slovenica. 2009; 94: 95-102.
- 40. Sankar S., Philip B., Philip R., Singh I.S.B. Effect of probiotics on digestive enzyme activities and growth of cichlids, Etroplus suratenis (Pearl spot) and *Oreochromis mossambicus* (Tilapia). Aquaculture Nutririon Journal. 2007; 23: 852-864.
- 41. Jang I.S., Ko Y.H., Kang S.Y., Lee C.Y. Effects of a commercial essential oil on growth performance, digestive enzyme activity and intestinal microflora population in broiler chickens. Animal Feed Science and Technology. 2007; 134: 304-315.
- 42. Boehm T. Evolution of vertebrate immunity. Current Biology. 2012; 22: 722-732.
- 43. Meng X., Cai H., Li H., You F., Jiang A., Hu W., Zhou Z. Clostridium butyricum-fermented Chinese herbal medicine enhances the immunity by modulating the intestinal microflora of largemouth bass (*Micropterus salmoides*). Aquaculture. 2023; 562: 738768.
- 44. Valenzuela-Gutiérrez R., Lago-Lestón A., Vargas-Albores F., Cicala F., Martínez-Porchas M. Exploring the garlic (*Allium sativum*) properties for fish aquaculture. Fish Physiology and Biochemistry. 2012; 47:1179-1198.
- 45. Yilmaz S., Ergün S. Effects of garlic and ginger oils on hematological and biochemical parameters of sea bass, *Dicentrarchus labrax*. Journal Aquatic Animal Health. 2012; 24: 219-224.
- 46. Immanuel G., Uma R.P., Iyapparay P., Citarasu T., Punitha S.M., Peter M., Michael B., Palavesam A. Dietary medicinal plant extracts improve growth, immune-activity and survival of tilapia *Oreochromis mosssambicus*. Journal of Fish Biology. 2009; 74: 1462.
- 47. Shalaby A. M., Khattab Y.A., Abdel Rahman A.M. Effects of garlic (*Allium sativum*) and chloramphenicol on growth performance, physiological parameters and survival of Nile Tilapia immunostimulants for fish. Journal of Ethnopharmacology. 2006; 88: 99-106.
- 48. Binaii M., Ghiasi M., Farabi S.M., Pourgholam R., Fazli H., Safari R., Alavi S.E., Taghavi M.J., Bankehsaz Z. Biochemical and haemato-immunological parameters in juvenile beluga (*Huso huso*) following the diet supplemented with nettle (*Urtica dioica*). Fish Shellfish Immunology. 2014; 36: 46-51.
- 49. Christybapita D., Divyagnaneswari M., Michael R.D. Oral administration of *Eclipta alba* leaf aqueous extract enhances the non-specific immune responses and disease resistance of *Oreochromis mossambicus*. Fish and Shellfish Immunology. 2007; 4:840-852.
- 50. Sarhadi I., Alizadeh E., Ahmadifar E., Adineh H., Dawood M.A. Skin Mucosal, Serum Immunity and Antioxidant Capacity of Common Carp (*Cyprinus sp.*) Fed Artemisia (*Artemisia annua*). Annals of Animal Science. 2020; 20: 1011-1027.
- 51. Magnadóttir B. Innate immunity of fish (overview). Fish and Shell fish Immunology. 2006; 20: 37–151.
- 52. Biller-Takahashi J.D., Takahashi L., Saita M., Gimbo R.Y., Urbinati E.C. Leukocytes respiratory burst activity as indicator of innate immunity of pacu *Piaractus mesopotamicus*. Brazi Journal Biology. 2013; 73: 425-429.
- 53. He J., Qiang J., Gabriel N.N., Xu P., Yang R. Effect of feeding-intensity stress on biochemical and hematological indices of GIFT tilapia, *Oreochromis niloticus*. Turkish Journal of Fisheries and Aquatic Sciences. 2015; 15: 305-312.
- 54. Livingstone D.R. Contaminant-stimulated reactive oxygen species production and oxidative damage in aquatic organisms. Marine Pollution Bulletin. 2001; 42: 656-666.
- 55. Ritola O., Tossavainen K., Kiuru T., Lindstorm-Seppa P., Molsa H. Effect of continuous and episodic hyperoxia on stress and hepatic glutathione levels in one-summer old rainbow trout, *Oncorhynchus mykiss*. Journal of Applied Ichthyology. 2002; 18: 159-164.
- 56. Manno M., Bertazzon A., Burlina A., Galzigna L. Interaction of low doses of ionizing radiation and carbon tetrachloride on liver superoxide dismutase and glutathione peroxidase in mice. Enzyme. 1985; 34: 107-112.
- 57. Metwally M.A.A. Effects of garlic, *Allium sativum* on some antioxidant activities in Tilapia, *Oreochromis niloticus*. Fisheries Marine Science Journal. 2009; 1: 56-64.

- 58. Islam M.N., Amel M. El., Amany A.A. Influence of dietary ginger (*Zingiber officinale*) on haemato-biochemical parameters, spleen histology and resistance of *Oreochromis niloticus* fingerlings to *Aeromonas hydrophila* infection. Egypt Journal of Aquatic. 2016; 6: 25-45.
- 59. Saha N.A., Usami T., Suzuki Y. Seasonal changes in the immune activities of common carp (*Cyprinus* carpio). Fish Physiology and Biochemistry. 2002; 26: 379-38.
- 60. Ahmadifar E., Hoseinifar S.H., Adineh H., Moghadam M.S., Dawood M.A. Assessing the impact of purslane (*Portulaca oleracea* L.) on growth performance, anti-oxidative, and immune activities in grass carp (*Ctenopharyngodon idella*). Annals of Animal Science. 2020; 20: 1427-1440.
- 61. Yagi K. Assay for plasma lipid peroxidase. Methods in Enzymology. 1984; 109: 328-331.
- 62. Giannenas I., El-Triantafillou S., Stavrakakis M., Margroni S., Mavridis T., Steiner E. Assessment of dietary supplementation with carvacrol or thymol containing feed additives on performance, intestinal microbiota and antioxidant status of rainbow trout (*Oncorhynchus mykiss*). Aquaculture. 2012; 350-353.
- 63. Jafarinejad R., Gharaei A., Mirdar Harijani J. Dietary ginger improve growth performance, blood parameters, antioxidant capacity and gene expression in *Cyprinus carpio*. Iranian Journal of Fisheries Sciences. 2020; 19: 1237-1252.