

Original Article

Effect of Solvent Type on Rosmarinic Acid, Total Phenol, Flavonoids, and Antioxidant Activity of *Nepeta asterotricha* Rech. f: An Endemic Plant from Iran

Mohammad Hossein Mirjalili¹, Ali Sonboli², Atousa Aliahmadi² and Seyed Mostafa Goldansaz^{3*}

¹Department of Agriculture, Medicinal Plants and Drugs Research Institute, Shahid Beheshti University, Tehran, Iran ²Department of Biology, Medicinal Plants and Drugs Research Institute, Shahid Beheshti University, Tehran, Iran ³Department of Biology, Medicinal Plants and Drugs Research Institute, Shahid Beheshti University, Tehran, (Mahan Golsay Company, Yazd) Iran

Article History: Received: 31 July 2020/Accepted in revised form: 30 September 2020 © 2012 Iranian Society of Medicinal Plants. All rights reserved.

Abstract

The current study aims to investigate the effect of solvent variety on some phytochemical factors of *Nepeta asterotricha* Rech. f.: as an endemic plant from Iran. It is suspected that quantity and quality of chemical compounds would be affected by solvent type. For this purpose, aqueous, hydro-alcohol (50/50 ethanol/water), and methanol were used as solvents. The studied factors were total phenol, flavonoid, antioxidant activity, and rosmarinic acid that were measured by Folin-Ciocalteu, colorimetric, FRAP, and HPLC methods, respectively. The outcomes showed hydro-alcohol extraction could significantly isolate phenol compounds (156.84 mg GAE/g DW) with considerable antioxidant properties (318.55 mg AA/g DW). However, methanol was more effective to extract total flavonoid content (101.34 mg RE/g DW) and rosmarinic acid (140.39% DW). In addition, more study should be done to investigate the importance of each compounds for both medicinal and industrial uses.

Keywords: Folin-Ciocalteu, Colorimetric, FRAP, HPLC, Endemic.

Introduction

Nepeta L. is demonstrated by 79 species in the Iranian flora, 42 of which are endemic such as *Nepeta asterotricha* Rech. f. [1,2]. The species has been used as febrifuge, anti-septic, antitussive, diaphoretic, sedative, anti-asthmatic, antipyretics, against snakes and scorpion bites, feline and canine attractants [3,4]. Other therapeutic effects are antioxidant [5,6], cytotoxic [7], Antifungal [8], and anti-inflammatory [9].

Medicinal plants are valuable for their therapeutic effects that have chemical components serving as natural products, or secondary metabolites such as phenol, flavonoids, organic acids, terpenes, terpenoids, among others. In other words, these compounds can be used for human disease treatment [10-18]. Characterization and isolation of compounds would be influenced by the extraction methods and condition. In addition, variation in composition and pharmacologic properties are related to technical practices among different laboratories briefly named as post-harvest techniques such as solvent type, sample preparation, and extraction method [19-22].

The present study aims to investigate total phenolic compounds, antioxidant activity, and variation the content of rosmarinic acid from *N. asterotricha* Rech. f. in different solvent. For this purpose, ethanol, methanol, and aqueous were used to extract aerial part of the genus. It is noteworthy that the current research consists of two main objective, which the first one is influence of solvent on the studied phytochemical parameters in *N. asterotricha* Rech. f. The other one is evaluation of the effect of solvents on antioxidant activity and the amount of rosmarinic acid.

Method and Material

Plant Material

*Corresponding author: Department of Biology, Medicinal Plants and Drugs Research Institute, Shahid Beheshti University, Tehran, (Mahan Golsay Company, Yazd) Iran Email Address: mostafagoldansaz@yahoo.com The plant materials were prepared at full flowering stage in Deh-Bala, Yazd, Iran. The sample were identified in Herbarium of Medicinal Plants and Drugs Research Institute, Shahid Beheshti University, Tehran, Iran. The Herbarium code of *N. asterotricha* Rech. f. is MPH-2569.

Plant Extraction

In order to formulate different extract, several solvents were prepared for maceration method. The studied solvents were methanol (MeOH), water (aqueous), and hydro-alcohol (50% water: 50% EtOH). Each solvent was separately added to dried material of the aerial part of *N. asterotricha* with 20 times ratio. Then, the new mixtures were shaken for 24 hours and were separately filtered. Each sample was extracted two times. Finally, the solvents were completely removed and three types of extraction were kept in shade for further investigation.

Total Phenol Content

Folin–Cicalteu reagent was used to determine the concentration of total phenolic compounds [22]. Briefly, 5 μ l of each extract (2 mg/ml), 195.5 μ l of distilled water, and 12.4 μ l of Folin–Cicalteu was mixed to microplate wells. After 3 minutes, 37.15 μ l of 7% (w/v) sodium carbonate was added, too. The newly obtained solution was shaken for 120 minutes and absorbance was measured at 765 nm. Finally, based on standard Gallic acid (GA) curve, total phenol compounds were calculated and expressed as mg of Gallic acid equivalents (GAE) per g of dry weight (DW).

Total Flavonoid Content

According to colorimetric method, total flavonoid content was measured. In detail, 20 µl of each extract (10 mg/ml) was mixed with 80 µl of distilled water and 6 µl of NaNO₂ (15%) and allowed to stand for 6 minutes. Then, 6 µl of 10% AlCl₃ solution was added. After 6 minutes, 80 µl of 4% NaOH and 8 µl distilled water was added to the solution. The obtained mixture was shaken for an hour and absorbance was measured at 510 nm. The total flavonoid content of the different extracts was expressed as mg of rutin equivalents per g of dry weight of plant material (mg RE/g DW).

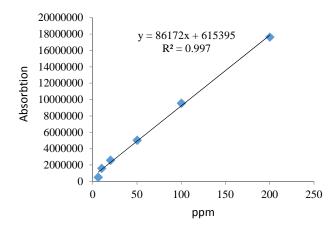
Antioxidant Activity

The ferric reducing antioxidant power (FRAP) was carried out using standard method. In order to prepare FRAP reagent, 300 mmol sodium acetate buffer (pH 3.6), 20 mmol iron (III) chloride solution, and 10 mmol TPTZ solution in 40 mmol HCl in a volume ratio of 10:1:1 were mixed. It is worth to note that prepared FRAP have to use fresh that is stable for 30 minutes. After that, 200 μ L of FRAP was added to 20 μ L of each extract (0.5 mg/mL). Then it was incubated at 37 °C for 30 min and the absorbance was determined at 593 nm. Different

concentrations of iron (II) sulfate solution (0.0625-1 mmol) were prepared to do standard solutions for the calibration curve. It should be mentioned that ascorbic acid was used as control sample and the results were expressed in mg ascorbic acid per g of dry weight extract.

Rosmarinic Acid in the Plant Extracts

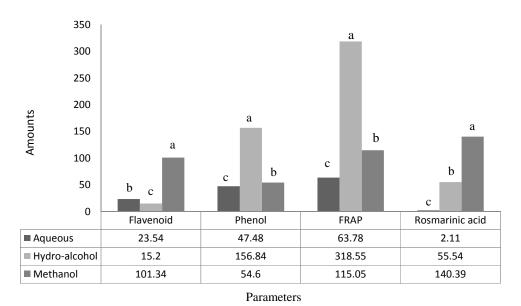
Isolation of rosmarinic acid was performed using HPLC. A waters liquid chromatography apparatus consisted of a Separations module: waters 2695 (USA) and Dual absorbance Detector waters 2487 (USA) were used for the HPLC analysis. Injection was auto sampler injector equipped with a 100 µl loop. Data acquisition and integration was performed with Millennium32 software. The chromatographic assay was performed on a 25 cm×4.6 mm with pre-column, Eurospher 100-5 C₁₈ analytical column provided by KNAUER (Berline, Germany) reversed phase matrix (5 µm) (Waters) and elution was carried out in a gradient system with acetonitrile as the organic phase (solvent A) and distilled water (solvent B) with the flow-rate of 1 mL min⁻¹. Peaks were monitored at 330 nm wavelength. Injection volume was 20 µL and the temperature was maintained at 25°C. Calibration graphs (figure 1) were plotted subsequently for linear regression analysis of the peak area with concentration 6, 10, 20, 50, 100, and 200 mg L⁻¹.





Statistical Analysis

In order to analyze the data, all of them were subjected to SAS version 9.4. Probability levels of 1% and 5% (P < 0.01 or 0.05) were used to test the significance among the treatments. All analyses were conducted in three independent replicates. The mean values and standard deviations for all studied factors were calculated and the obtained outcomes were expressed as mean \pm SD. Least significant difference (LSD) was used to split the means of main effect, when an F-test showed statistical significance.



Results and Discussion

In order to extract phytochemical compounds, solvent type is one of the most important factor [23]. For this purpose, various solvents (water and organic solvent) with different polarities were used to comprise the solvent effect. The TPC, TFC, antioxidant activity, and rosmarinic acid of the studied solvent were in the range of 54.6-156.84 mg GAE/g DW, 15.2-101.34 mg RE/g DW, 63.78-318.55 mg AA/g DW, and 2.11-140.39% DW, respectively. There is noteworthy that high level of TPC in extract does not cause a high amount of TFC. The present results showed hydro-alcohol extract could isolate more phenol compounds, significantly. Use of ethanol as a polar organic solvent could cause this result and it is in accordance with other research that revealed combination of aqueous organic solvent is more useful than absolute organic solvent to extract TPC [24]. However, some researchers believed that the amount of TPC could be increased using ethanol as an absolute organic solvent [25,26]. Comparing the effect of the studied solvents showed highly significant difference for the total flavonoids. The present outcome showed methanol was the best solvent to extract the total flavonoids content and it was similar to previous study [27].

Fig. 2 The effect of solvent type on the studied parameters

The antioxidant properties of hydro-alcohol extraction was the best one. This activity is associated with the high level of phenol compounds, which is in accordance with previous studies [18,19,28,29]. On the other hand, the present results like other study emphasized aqueousorganic solvents verified higher free radical scavenging activity than their absolute organic preparations [24]. Despite of these outcomes, some research showed the radical scavengers were more soluble in absolute organic solvent than aqueous one [30]. Based on these variations, it is speculated the type of solvent, polarities, and the existence of polyphenolic compounds have effect on the antioxidant activities.

One of the most important phenolic acids is rosmarinic acid that is an ester of caffeic acid and 3.4dihydroxyphenyllactic acid derived from hydroxycinnamic acid [31]. This compound showed several biological properties such as antioxidant and antimutagen [32], hepatoprotective [33], anti-acetyl cholinesterase properties [34], neuroprotective [35], antiviral, antibacterial, and anti-inflammatory [36,37], so investigation on this compound is valuable. Use of methanol as a solvent can obtained the highest amount of rosmarinic acid. In addition, the current research like other researches showed the phytochemical compositions have been significantly affected by solvent [20,23].

Acknowledgment

This research was supported by Shahid Beheshti University that is gratefully appreciated.

References

- 1. Jamzad Z. A survey of Lamiaceae in the flora of Iran. Rostaniha. 2013;14:59-67.
- 2. Mozaffarian V. Identification of medicinal and aromatic plants of Iran, Farhang Moaser. 2013.

- 3. Amin G. Popular medicinal plants of Iran, Iranian Ministry of Health Publications. 1991.
- 4. Zargari A. Medicinal plants (4th ed), Tehran University. 1995.
- Duda S.C, M rghita L.A, Dezmirean D, Duda M, M rg oan R, Bobi O. Changes in major bioactive compounds with antioxidant activity of *Agastache foeniculum*, *Lavandula angustifolia*, *Melissa officinalis* and *Nepeta cataria*: Effect of harvest time and plant species. Ind Crops Prod. 2015;77:499-507.
- Sarikurkcu C, Ceylan O, Targan S, avar Zeljkovi S. Chemical composition and biological activities of the essential oils of two endemic *Nepeta* species. Ind Crops Prod. 2018;125:5-8.
- Skori M, Gligorijevi N, avi M, Todorovi S, Jankovi R, Risti M, Radulovi S. Cytotoxic activity of *Nepeta rtanjensis* Dikli & amp; Milojevi essential oil and its mode of action. Ind Crops Prod. 2017;100:163-170.
- Kumar V, Mathela C.S, Tewari G, Singh D. Antifungal activity of *Nepeta elliptica* Royle ex Benth. oil and its major constituent (7R)-trans,trans-nepetalactone: A comparative study. Ind Crops Prod. 2014;55:70-74.
- Goldansaz S.M, Festa C, Pagano E, De Marino S, Finamore C, Parisi O.A, Borrelli F, Sonboli A, D'Auria M.V. Phytochemical and biological studies of *Nepeta asterotricha* rech. f. (Lamiaceae): Isolation of Nepetamoside. Molecules. 2019;24:1684.
- 10. Mosleh Arani A, Naderi M, Goldansaz S.M. Effect of harvesting time on essential oil content and composition of *Thymbra spicata*. J of Med Plants and By-Prod. 2015;1:51-55.
- Dib I, Angenot L, Mihamou A, Ziyyat A, Tits M. Artemisia campestris L.: Ethnomedicinal, phytochemical and pharmacological review. J. Herb. Med. 2017;7:1-10.
- Goldansaz S.M, Hakimi Meybodi M, Mirhosseini A, Mirjalili M. Essential oil composition of *Salvia tebesana* Bunge (Lamiaceae) from Iran. Rec Nat Prod. 2017;11:310-314.
- Amiri A, Morakabati N. Encapsulation of *Satureja khuzestanica* Essential Oil in Chitosan Nanoparticles with Enhanced Antifungal Activity. Int J Food Sci Nutr. 2017;11:331-336.
- 14. Lemus-Mondaca R, Vega-Gálvez A, Rojas P, Stucken K, Delporte C, Valenzuela-Barra G, Pasten A. Antioxidant, antimicrobial and anti-inflammatory potential of *Stevia rebaudiana* leaves: effect of different drying methods. J Appl Res Med Aromat Plants. 2018;11:37-46.
- Singh D, Chaudhuri P.K. A review on phytochemical and pharmacological properties of Holy basil (*Ocimum sanctum* L.). Ind Crops Prod. 2018;118:367-382.
- 16. de Oliveira-Júnior R.G, Ferraz C.A, de Oliveira A.P, Araújo C.S, Oliveira L.F, da S, Picot L, Almeida, J.R.G, da S. Phytochemical and pharmacological aspects of *Cnidoscolus pohl* species: A systematic review. Phytomedicine. 2018;50:137-147.
- Goldansaz S.M, Jafarian Jeloudar Z, Safaeian R, Sonboli A. Comparison of the chemical constitutions, antibacterial, anti-Candida, and antioxidant activity of *Nepeta asterotricha* Rech. F. essential oil. American Journal of Essential Oils and Natural Products. 2019;7:15-22.
- Alara O.R, Abdurahman N.H, Ukaegbu C.I. Soxhlet extraction of phenolic compounds from *Vernonia cinerea* leaves and its antioxidant activity. J Appl Res Med Aromat Plants. 2018;11:12-17.

- Bampouli A, Kyriakopoulou K, Papaefstathiou G, Louli V, Krokida M, Magoulas K. Comparison of different extraction methods of *Pistacia lentiscus* var. *chia* leaves: Yield, antioxidant activity and essential oil chemical composition. J Appl Res Med Aromat Plants. 2014;1:81-91.
- 20. Amiri A, Mousakhani-Ganjeh A, Amiri Z, Guo Y, Singh A.P, Esmaeilzadeh Kenari R. Fabrication of cumin loaded-chitosan particles: Characterized by molecular, morphological, thermal, antioxidant and anticancer properties as well as its utilization in food system. Food Chem. 2020;310:125821.
- Oreopoulou A, Papavassilopoulou E, Bardouki H, Vamvakias M, Bimpilas A, Oreopoulou V. Antioxidant recovery from hydrodistillation residues of selected Lamiaceae species by alkaline extraction. J Appl Res Med Aromat Plants. 2018;8:83-89.
- Javanmardi J, Stushnoff C, Locke E, Vivanco J. Antioxidant activity and total phenolic content of Iranian Ocimum accessions. Food Chem. 2003;83:547-550.
- Dorta E, Lobo M.G, Gonzalez M. Reutilization of Mango byproducts: study of the effect of extraction solvent and temperature on their antioxidant properties. J Food Sci. 2012;77:80-88.
- 24. Bhebhe M, Füller T.N, Chipurura B, Muchuweti M. Effect of solvent type on total phenolic content and free radical scavenging activity of black tea and herbal infusions. Food Anal Methods. 2016;9:1060-1067.
- 25. Figueroa-Espinoza M.C, Zafimahova A, Alvarado P.G.M, Dubreucq E, Poncet-Legrand C. Grape seed and apple tannins: Emulsifying and antioxidant properties. Food Chem. 2015;178:38-44.
- 26. Hwang E.S, Thi. Effects of extraction and processing methods on antioxidant compound contents and radical scavenging activities of Laver (*Porphyra tenera*). Prev Nutr Food Sci. 2014;19:40-48.
- Elfalleh W, Kirkan B, Sarikurkcu C. Antioxidant potential and phenolic composition of extracts from *Stachys tmolea*: An endemic plant from Turkey. Ind Crops Prod. 2019;127:212-216.
- Burkhardt A, Sintim H.Y, Gawde A, Cantrell C.L, Astatkie T, Zheljazkov V.D, Schlegel V. Method for attaining fennel (*Foeniculum vulgare* Mill.) seed oil fractions with different composition and antioxidant capacity. J Appl Res Med Aromat Plants. 2015;2:87-91.
- 29. Dincer C, Torun M, Tontul I, Topuz A, Sahin-Nadeem H, Gokturk R.S, Ozdemir F. Phenolic composition and antioxidant activity of *Sideritis lycia* and *Sideritis libanotica* subsp. linearis: Effects of cultivation, year and storage. J Appl Res Med Aromat Plants. 2017;5:26-32.
- 30. Rigane G, Ghazghazi H, Aouadhi C, Ben Salem R, Nasr Z. Phenolic content, antioxidant capacity and antimicrobial activity of leaf extracts from *Pistacia atlantica*. NAT PROD RES. 2017;31:696-699.
- 31. Lu Y, Yeap Foo L. Polyphenolics of *Salvia*-a review. Phytochemistry. 2002;59:117-140.
- 32. Petersen M, Simmonds M.S. Rosmarinic acid. Phytochemistry. 2003;62:121-125.
- 33. Renzulli C, Galvano F, Pierdomenico L, Speroni E, Guerra M.C. Effects of rosmarinic acid against aflatoxin B1 and ochratoxin-A-induced cell damage in a human hepatoma cell line (Hep G2). J Appl Toxicol. 2004;24:289-296.
- Falé P.L, Borges C, Madeira P.J.A, Ascensão L, Araújo M.E.M, Florêncio M.H, Serralheiro M.L. M. Rosmarinic acid,

scutellarein 4 -methyl ether 7-O-glucuronide and (16S)-coleon E are the main compounds responsible for the antiacetylcholinesterase and antioxidant activity in herbal tea of *Plectranthus barbatus* ("falso boldo"). Food Chem. 2009;114:798-805.

- 35. Fallarini S, Miglio G, Paoletti T, Minassi A, Amoruso A, Bardelli C, Lombardi G. Clovamide and rosmarinic acid induce neuroprotective effects in *in vitro* models of neuronal death. Br J Pharmacol. 2009;157:1072-1084.
- 36. Osakabe N, Yasuda A, Natsume M, Yoshikawa T. Rosmarinic acid inhibits epidermal inflammatory responses: anticarcinogenic effect of *Perilla frutescens* extract in the murine two-stage skin model. Carcinogenesis. 2003;25:549-557.
- 37. Swarup V, Ghosh J, Ghosh S, Saxena A, Basu A. Antiviral and anti-inflammatory effects of rosmarinic acid in an experimental murine model of *Japanese encephalitis*. Antimicrob. Agents Chemother. 2007;51:3367-3370.