

The Anti-Cancer Arsenal of the Date Palm (*Phoenix dactylifera* L.): A Review of Bioactive Pathways Across Various Organs

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Article Info	ABSTRACT
Article Type Original Article Article History Received: 12 July 2025 Accepted: 18 September 2025 © 2012 Iranian Society of Medicinal Plants. All rights reserved. *Corresponding author ztoluei@kashanu.ac.ir 	The utilization of natural plant compounds for medicinal applications plays a crucial role in cancer treatment. Cancer arises through various mechanisms, and the date palm (<i>Phoenix dactylifera</i>) is notable for its rich array of anticancer compounds found in different organs. This review evaluates the anticancer mechanisms of various organs of the date palm, emphasizing the contribution of their shared and unique compounds. This study, was extracted and reviewed all scientific books and articles related to the anticancer properties of date palm seeds, roots, hearts, leaves, inflorescences (spathe, flowers, and pollen), and fruits from online databases using specialized keywords from 1980 to 2025. The date palm minerals and bioactive compounds may not only aid in cancer prevention but also slow its progression, enhance the efficacy of chemotherapeutic agents, and improve overall treatment outcomes. The bioactive components derived from the date palm exhibit significant abilities to inhibit cancer cell proliferation and induce apoptosis. Although previous research has largely studied the fruit, there is a growing need to study the bioactive compounds of other organs. Findings confirm that underrepresented organs offer potent anticancer bioactivities. Concentrating on optimizing these potent compounds, particularly from less-studied parts, may yield promising results. Future investigations should focus on isolating compounds from various date palm organs, which exhibit varied phytochemical profiles across cultivars. This approach may aid in harnessing the therapeutic potential of the date palm against cancer. Keywords: Anticancer, Arecaceae, Bioactive components, Minerals

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INTRODUCTION

Cancer is considered one of the most complex and incurable diseases of the past few centuries [1]. Recently, there has been a growing interest in utilizing natural medicinal plants as affordable alternatives for cancer treatment [2]. The date palm (*Phoenix dactylifera* L.) is a tropical plant from the Arecaceae Bercht. & J.Presl family, is commonly found in the countries of the Persian Gulf and tropical regions of the Northern Hemisphere [3]. The date palm exhibits a complex root system characterized by a network of fibrous roots that facilitate nutrient uptake and provide stability in sandy soils. Palms possess long, pinnate, or fan-shaped leaves that emerge from a central crown, playing a crucial role in photosynthesis and transpiration. The date palm seed is a hard, elongated structure encased in a fleshy mesocarp, functioning as a reproductive unit that can germinate under favorable conditions. The date palm produces intricate, branched inflorescences encased in a protective spathe, with pollen grains that are vital for the fertilization of flowers, leading to fruit development. The palm's apical meristem, or heart, is an essential growth point that gives rise to new leaves and plays a critical role in the overall health and vigor of the plant. The date fruit is a single, one-seeded berry, oblong, terete, with a terminal stigma, a fleshy pericarp, and a membranous endocarp present between the seed and the flesh [4, 5]. Iran is the world's fourth-largest producer of dates, and its date production is expected to increase by 20.55% by 2028 [6]. There are around 3000 different date palm cultivars in the world, each with varying contents of salts, flavonoids, phenolic compounds, and fatty acids

in their organs [7]. Hypotheses have been proposed about date palm organs' anticancer and antitumor properties [8]. Over the last four decades, there has been extensive research on compounds found in dates. The cancer suppressor pathways reviewed in this article are shown in Figure 1. The present study was conducted to provide a comprehensive review of the anticancer potential of different organs of the date palm, focusing on the shared presence of minerals and bioactive constituents.

MATERIALS AND METHODS

This study systematically collected and reviewed scientific books and research articles addressing the anticancer properties of *P. dactylifera* organs, including seeds, roots, hearts, leaves, inflorescences (spathe, flowers, and pollen), and fruits. Relevant literature was retrieved from major online databases such as NCBI, SpringerLink, PubMed, and Google Scholar, covering the period from 1980 to 2025. The search employed specialized keyword combinations, including anticancer or antioxidant potential/effect + date palm [organ], minerals/bioactive compounds + date palm [organ], and anticancer pathways + bioactive compounds/minerals, to ensure the comprehensive inclusion of studies reporting the biological activities and chemical constituents of different date palm organs. In this article, an attempt has been made to discuss the most striking anticancer compounds in the section related to each organ, and if those compounds exist in other organs, they are listed in tables.

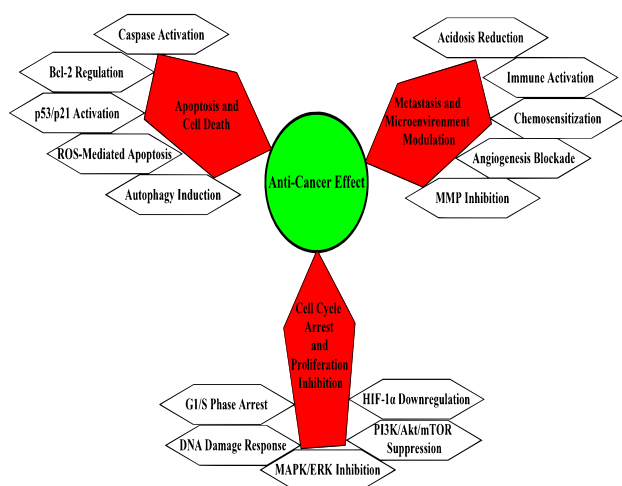


Fig. 1 Pathways of anticancer effects associated with date palm

Ethnobotanical, Nutritional, and Pharmacological Applications of Date Palm Organs

The date palm is repeatedly mentioned in the Holy Qur'an as a plant of blessing and sustenance, and its multiple organs have been integrated for centuries into traditional diets, medicine, and crafts [9]. Ethnobotanical records show that the roots are used in decoctions to relieve toothache, stomach upset, and intestinal disorders, while modern studies have detected phenolic acids and antioxidant compounds in root extracts that explain their antimicrobial and anti-inflammatory actions [10-12]. In comparative analyses, leaves contain higher total phenolic and flavonoid contents than roots, conferring stronger radical-scavenging and anti-inflammatory properties [12]. Traditionally, leaves are boiled for treating hypertension and fever, and the dried fronds are woven into mats, baskets, and handicrafts, illustrating both medicinal and practical utility [11]. The pollen, consumed to improve male infertility and sexual weakness, has shown a significant increase in serum testosterone and a reduction in luteinizing hormone in controlled human trials, confirming its long-standing role in reproductive medicine [13]. The seeds, rich in polyphenols and natural antioxidants, are ground into powders to aid liver disorders and diabetes, while experimental studies highlight their antimicrobial and cytoprotective effects that support nutraceutical development [14]. The heart of the palm, tender and nutrient-dense, is eaten raw in salads and incorporated into functional foods, such as low-fat camel-milk ice cream, where it enhances protein and fiber content and maintains microbial stability [15]. Comparative analyses demonstrated that the male flowers possess greater phenolic and flavonoid concentrations than the spathe, explaining their stronger antioxidant capacity and justifying their use in herbal tonics for fatigue, urinary infections,

and general weakness [16]. Collectively, these findings validate the traditional and medicinal importance of each organ, confirming *P. dactylifera* as a culturally, nutritionally, and pharmacologically significant species.

Anticancer Properties of Date Seeds

The compounds in date palm seeds vary depending on the palm cultivar [17]. Date seeds contain varying amounts of carbohydrates, minerals, fats, water, phenolic compounds, flavonoid compounds, and antioxidants. Two of the main phenolic compounds in these seeds are vanillin and catechin [18]. Vanillin is associated with vanilloid receptors and has been found to induce changes in cancer cells. These changes include decreasing the levels of cyclin D1/A2/B, p38, and p65, while also increasing the levels of p53 and p21 [19]. This suggests that vanillin could be effective in treating cancer. Catechin inhibits the growth of cancer cells and can make them undergo apoptosis by increasing the activity of III, VIII, and IX caspases and p38 phosphorylation, and regulating the expression of cyclins A and B1 [20-22]. Flavonoids and Non-flavonoid phenolic compounds of different organs have been reviewed in Table 1 and Table 2, respectively. The Date seed oil contains four major phenolic compounds, including 2-hydroxycinnamic acid, vanillic acid, 4-hydroxybenzoic acid, and 4-hydroxycinnamic acid [23]. 2-Hydroxycinnamic acid can increase the expression of caspase III and annexin V, aiding in destroying the DNA of cancer cells [24]. Vanillic acid has the potential to suppress HIF-1 α expression, halting cancer cells in the G1 phase [25, 26]. It may also have anticancer effects in obese cancer patients [27]. 4-Hydroxybenzoic acid inhibits HSP90 α by aiding in its acetylation, reducing cancer cell resistance to Adriamycin by inhibiting the histone deacetylase 6 enzyme, thereby decreasing cancer cell survival [28-30]. 4-Hydroxycinnamic acid can also reduce cancer cell proliferation and has potential use as a vector in targeted cancer treatment, while also preventing oxidative stress [31, 32]. Types of tocopherols have been found in date seeds [33]. The most abundant type in these seeds is alpha-tocopherol, which is important in cancer prevention, probably with its antioxidant activity [34]. Oleic acid and Lauric acid are the main fatty acids found in date seed extract (Table 3) [35, 36]. Oleic acid can prevent cancer cell proliferation by stopping the cell cycle in the G1 phase, while lauric acid induces cancer cell apoptosis by increasing reactive oxygen levels [37, 38]. The bioactive compounds in date seed extract can remain active in the digestive system [39, 40]. This extract significantly reduces the proliferation of liver, colorectal, and breast cancer cells through apoptosis mechanisms [41]. Date seeds protect against oxidative stress by reducing enzymes like lactate dehydrogenase and creatine kinase [42]. In general, depending on the palm cultivar, date seeds may have a potential impact on cancer treatment [43, 44].

Table 1 Flavonoids and anti-cancer mechanisms in different organs of the date palm.

Compound	Anticancer mechanism	Found in	References
Catechin	↑ Caspases III/VIII/IX; Regulates cyclins A/B1 → Apoptosis	Fruit, Heart, Leaf, Pollen, Seed	[18, 20-22, 58, 69, 92, 96, 120]
Quercetin	Activates caspase III → Apoptosis/autophagy	Fruit, Heart, Leaf, Pollen	[58, 69, 92, 96, 98, 120, 122, 125]
Luteolin	Induces cell cycle arrest via DNA damage/redox regulation	Fruit, Leaf, Seed	[18, 94, 98, 120, 123, 125]
Apigenin	↓ PI3K, MMP; ↑ Bax, p53, p16	Fruit, Heart, Leaf, Pollen	[58, 69, 94, 96, 98, 120, 124, 125]
Naringenin	↓ ERK1/2/JNK phosphorylation; ↑ Caspase III, p53 → Apoptosis (melanoma)	Fruit, Heart, Pollen	[58, 62, 63, 94, 96, 98, 125]
Isorhamnetin	Inhibits PI3K/AKT signaling → Antitumor effects	Pollen, Seed	[69, 94, 95]
Hesperidin	Activates caspases; Deactivates kinases → Apoptosis	Pollen	[96-98]
Rutin	Induces cell cycle arrest; ↓ DNA damage; ↑ Caspases, p53	Fruit, Heart, Leaf, Pollen	[58, 69, 92, 94, 96, 98, 125, 127]

Symbol Key: ↑ = Increase/Activation; ↓ = Decrease/Inhibition; → = Leads to.

Table 2 Non-flavonoid phenolic compounds and anti-cancer mechanisms in different organs of the date palm.

Compound	Anticancer mechanism	Found in	References
Vanillin	↓ Cyclin D1/A2/B, p38, p65; ↑ p53, p21	Heart, Seed	[18, 58, 69, 92, 96, 120]
2-Hydroxycinnamic acid	↑ Caspase III, Annexin V → DNA damage	Fruit, Pollen, Seed	[23, 24, 96, 120]
4-Hydroxybenzoic acid	Inhibits HSP90α acetylation → ↓ HDAC6 → ↓ Adriamycin resistance	Fruit, Root, Leaf, Seed	[12, 23, 28-30, 69, 120]
4-Hydroxycinnamic acid	↓ Cancer cell proliferation; Prevents oxidative stress	Fruit, Root, Leaf, Pollen, Seed	[12, 23, 31, 32, 69, 96, 120, 125]
Hydroxystilbenes	Inhibits Bcl-2; Activates caspases III/VII; Induces autophagy via Hsp-70 activation	Root	[45, 46]
Chlorogenic acid	↑ SUMO1 → ↓ miR-17; ↑ p21 → ↓ proliferation/invasion/motility	Heart, Pollen	[58, 59, 94, 96]
Syringic acid	Inhibits mTOR/AKT pathway; ↑ ROS → Mitochondrial membrane disruption	Fruit, Root, Leaf, Heart	[12, 49, 50, 58, 120]
Ferulic acid	↓ MMP-9 mRNA expression → ↑ chemotherapy efficacy	Fruit, Heart, Root, Leaf, Pollen, Seed	[12, 18, 51, 52, 58, 69, 94, 96, 120, 125]
Methyl gallate	↓ MMP9/MMP2; ↑ TIMP-2 → Inhibits migration/invasion	Heart, Leaf	[58, 69, 70]
Protocatechuic acid	↓ Ras/Akt/NF-κB pathway; Targets RhoB → Inhibits migration	Fruit, Leaf, Pollen	[69, 71, 72, 96, 120, 125]
3,4-Dimethoxy toluene	↑ GABA → Inhibits MMP-2/MMP-9; ↓ Gastric cancer/liver metastasis	Spathe	[72, 76-78, 80]
β-Caryophyllene	Binds CB2 receptor → ↑ Proapoptotic factors	Spathe	[78-80]
Caffeic acid	Triggers intrinsic apoptosis	Fruit, Heart, Pollen, Seed	[18, 23, 58, 92, 93, 96, 120]
Pyrogallol	Reduces tumor size (doxorubicin-like effect)	Pollen	[96, 99]
Ellagic acid	Regulates PI3K/Akt, JNK, mitochondrial, Bcl-2/Bax, TGF-β/Smad3 pathways	Heart, Pollen	[58, 96, 98, 100]
Quinic acid	↓ Cyclin D1 → Inhibits oral cancer proliferation	Fruit	[125, 126]
Vanillic acid	Suppresses HIF-1α → G1 phase arrest	Fruit, Pollen, Seed	[18, 23, 25-27, 92, 96, 120]
Epicatechin	↑ Sensitivity to radiation/chemotherapy	Fruit, Leaf, Pollen, Seed	[69, 92, 96, 120, 127, 128]

Symbol Key: ↑ = Increase/Activation; ↓ = Decrease/Inhibition; → = Leads to.

Table 3 Common non-phenolic substances and anticancer mechanisms in different organs of date palm.

Substance	Classification	Anticancer mechanism	Found in	References
Alpha-tocopherol	as Vitamin E	Prevents cancer by its antioxidant activity	Seed, Pollen, Fruit	[18, 33, 34, 101, 121]
Lauric acid	Fatty acid	Induces apoptosis by increasing ROS	Seed, Pollen	[18, 23, 33, 35, 38, 98, 101]
Lysine	Amino acid	Reduces metastasis and enhances chemotherapy	Pollen, Fruit	[101, 106-109]
Oleic acid	Fatty acid	Arrests cell cycle in G1 phase	Seed, Pollen	[18, 23, 33, 35-37, 98, 101]
Palmitic acid	Fatty acid	Upregulates caspases and p53, suppresses Bcl-2	Seed, Pollen	[18, 23, 33, 35, 98, 101-103]
Phenylalanine	Amino acid	Serves as a drug carrier and inhibits prostate cancer	Pollen, Fruit	[101, 106, 110, 111]

Anticancer Properties of Date Tree Roots

Date palm roots contain significant amounts of Hydroxystilbenes [45]; Stilbenes can enter cancer cells to undergo apoptosis by inhibiting anti-apoptotic proteins such as Bcl-2 and activating III and VII caspases. They can also force cancer cells to undergo autophagy by activating Hsp-70. They can also initiate antioxidant cascade pathways by activating PI3K/Akt [46]. One of the highest elements absorbed by date palm roots is potassium [47]. Elevated levels of potassium have been shown to correlate with reduced cancer risk and the suppression of tumor growth; This is due to the role of potassium in regulating cell growth and proliferation [48]; In Table 4, the minerals of different organs have been reviewed. Syringic acid and ferulic acid are two notable phenolic compounds in date palm roots [12]. Syringic acid may exhibit notable anticancer properties by modulating crucial signaling pathways that affect oncogenic transcription factors and apoptosis-related proteins, leading to the inhibition of proliferation across various cancer types, such as colorectal and breast cancer. This compound can regulate cell cycle progression and promote apoptosis by upregulating pro-apoptotic proteins while downregulating anti-

apoptotic factors, potentially enhancing the effects of conventional chemotherapy through synergistic actions targeting pathways related to cancer resistance and progression. Additionally, syringic acid may disrupt mitochondrial membrane potential by increasing the generation of reactive oxygen species (ROS) and can inhibit the mTOR/AKT signaling pathway, thus representing a promising candidate for therapeutic intervention in gastric cancer through its complex anti-cancer mechanisms [49, 50]. Ferulic acid prevents cancer cell proliferation, induces apoptosis by reducing MMP-9 mRNA expression, and enhances the effectiveness of anticancer drugs in chemotherapy [51, 52]. *Geotrichum candidum* and *Penicillium citrinum* are two fungal endophytes found in date palm roots [53]. It has been shown that the extracts of these endophytes have significant antioxidant properties, and it is speculated that unknown metabolites extracted from them can be effective in slowing down the progression of cancer. Although the research about these roots is limited, the fact that silver nanoparticles synthesized from them have shown anti-cancer activity can be promising to focus more on this organ in the future [54].

Table 4 Minerals and a summary of their anti-cancer effects in different organs of the date palm.

Minerals	Anti-cancer effects	Found in	References
Potassium	Cell growth regulator, tumor suppression	Fruit, Root, Leaf, Seed, Heart, Pollen	[23, 35, 47, 48, 55, 65, 96, 106, 113, 121]
Indium	Caspase modulation, angiogenesis inhibition	Heart	[55, 56]
Strontium	ROS generation, apoptosis	Heart	[55, 57]
Zinc	DNA repair, caspase activation	Fruit, Seed, Leaf, Heart, Pollen	[55, 65, 66, 96, 101, 106, 121]
Manganese	Mitochondrial regulation, apoptosis	Fruit, Seed, Leaf, Heart, Pollen	[23, 55, 65, 67, 96, 101, 106, 121]
Selenium	Antioxidant, apoptosis inducer	Fruit, Seed, Leaf, Heart, Pollen	[55, 65, 101, 113, 114, 116]

Anticancer Properties of Date Palm Heart

In addition to the valuable minerals, the heart of the date palm also contains Indium and Strontium [55]. Indium (III) complexes can exhibit significant anticancer activity by targeting various signaling pathways involved in cell proliferation and survival. These complexes may induce apoptosis in cancer cells via the regulation of apoptotic markers such as caspases and Bcl-2 family proteins, potentially inhibiting tumor growth. Additionally, indium compounds can affect angiogenesis-related pathways, impairing the formation of new blood vessels supporting tumor growth. Moreover, the ability of Indium complexes to generate reactive oxygen species may synergistically enhance their cytotoxic effects against neoplastic cells [56]. Strontium complexes can exhibit significant anticancer potential by modulation of key cellular pathways. These complexes may increase reactive oxygen species (ROS) levels and decrease mitochondrial membrane potential (MMP), leading to cell cycle arrest and apoptosis in cancer cells, particularly in A375 melanoma cells. Such mechanisms indicate that strontium can enhance the selective cytotoxicity towards malignant cells while potentially preserving normal tissues [57]. The extract of date palm heart may significantly inhibit cell viability in a concentration-dependent manner, which can be attributed to alterations in cancer cell morphology and declining viability rates. The extract's cytotoxic effects align with its phenolic and flavonoid content, which may activate specific apoptotic pathways while inhibiting cancer cell proliferation; Chlorogenic acid and Naringenin are powerful phenolic compounds in this regard [58]. Chlorogenic acid may demonstrate anticancer properties through the modulation of critical signaling pathways that facilitate cancer cell differentiation. It can enhance the expression of SUMO1, leading to the sumoylation of c-Myc and the subsequent downregulation of the miR-17 family while promoting p21 expression. This signaling cascade can diminish cancer cell proliferation, invasion, and motility, ultimately fostering a differentiated state across various cancer cell types [59]. Moreover, in vivo experiments suggest that chlorogenic acid may inhibit tumor growth in hepatoma and glioma models, presenting a promising avenue for cancer therapy by instigating cellular education rather than direct cytotoxic effects. In addition, chlorogenic acid can be converted into dihydrocaffeic acid, which may exert significant anticancer effects through its engagement with vital cancer pathways. Dihydrocaffeic acid can induce selective apoptosis in specific cancer cell lines like MCF-7, PC-3, and HCT-116, thereby leading to decreased cell viability in a concentration-dependent manner. While its precise mechanisms remain unclear, it may act similarly to caffeic acid, influencing apoptosis and cell cycle regulation through key signaling molecules [60, 61]. Naringenin may exert notable anticancer effects by modulating multiple signaling pathways that govern tumor behavior and progression, particularly in melanoma and angiogenesis. It can inhibit cell proliferation and migration in melanoma cells, which may be associated with reduced phosphorylation of key proteins such as ERK1/2 and JNK MAPK, emphasizing its impact on the MAPK pathway. Furthermore, naringenin may induce apoptosis through the upregulation of pro-apoptotic factors like activated caspase III and p53 while lowering anti-apoptotic signals, thereby highlighting its potential role in therapeutic strategies against cancer [62, 63]. The extract of date palm heart may enhance the regulation of specific anti-cancer pathways, particularly through the elevation of antioxidant enzymes and suppression of apoptosis-related markers such as

caspase III and cyclooxygenase-2 levels. It is suggested that treatment with this extract might activate programmed cell death protein-1 (PD-1), which can help mitigate oxidative stress and associated cellular damage. Overall, the findings indicate that the heart of palm extract may serve as a valuable supplement for preventing oxidative injury in cancer therapies involving adriamycin [64].

Anticancer Properties of Date Leaves

Zinc and Manganese are among the elements found in date palm leaves [65]; Zinc may play a pivotal role in various anticancer pathways, potentially influencing the regulation of oxidative stress, DNA damage repair, cell cycle progression, and apoptosis. Key proteins involved include p53, caspases, copper/zinc superoxide dismutase, and various DNA repair proteins reliant on zinc for proper function. Zinc deficiency can disrupt these pathways, leading to increased DNA damage and cancer risk, while adequate zinc levels may enhance the structural stability of p53, activate caspases, and promote effective DNA repair mechanisms, contributing to its chemoprotective effects against malignancies such as colorectal, pancreatic, esophageal, and head and neck cancers [66]. Manganese in its divalent form, when complexed with arginine dithiocarbamate, has demonstrated significant anticancer potential by exhibiting cytotoxic effects against the MCF-7 breast cancer cell line. This complex may initiate apoptosis through various mechanisms, including the induction of G0/G1 phase cell cycle arrest, interference with DNA repair, and promotion of proteasomal inhibition. Specifically, the involved proteins and pathways include arginase, glutamine synthetase, Mn-superoxide dismutase, and the complex's interaction with DNA through hydrogen bonding, showcasing its multifaceted role in cancer therapy [67]. Date palm leaves contain important polyphenolic compounds and antioxidants [68]. Methyl gallate and protocatechuic acid are two significant compounds found in date palm leaves [69]. Methyl gallate inhibits cancer cell migration by decreasing MMP9 and MMP2 expression and increasing TIMP-2 expression and invasion without harming normal cells [70]. Protocatechuic acid stops cancer cell migration and induces apoptosis involving the down-regulation of Ras/Akt/NF- κ B pathway and MMP-2 production by targeting RhoB activation [71, 72]. Although the bioactive compounds in date palm leaves may vary depending on their cultivar, they still exhibit a positive effect in halting the cancer process [73, 74].

Anticancer Properties of Date Palm Inflorescences

The spathe

One of the main compounds of date spathe is 3,4-dimethoxy toluene [75]; This aromatic compound can increase the level of γ -amino butyric acid [76]; It can retard the proliferation rates and enhance apoptosis of leukemia cells, inhibit MMP-2 and MMP-9 activity and expression, decrease the number of gastric cancers of the glandular stomach and intrahepatic liver metastasis, and inhibit human liver cancer cell migration and invasion [77]. β -Caryophyllene oxide (in all) and β -Caryophyllene (in most) are common compounds in the spathe of different date cultivars [78]; β -Caryophyllene acts as a phytocannabinoid with a significant affinity for cannabinoid receptor type II but shows no binding to cannabinoid receptor type I, indicating that its mechanism of action may not involve the endocannabinoid system. β -Caryophyllene oxide can alter critical pathways involved in cancer development while decreasing pro-cancer gene expression and enhancing proapoptotic levels, suggesting that β -Caryophyllene

may serve as a potential natural analgesic and anticancer agent, especially valuable in improving the efficacy of traditional chemotherapy in cancer patients dealing with chronic pain [79]. Linalool, Carvacrol, and Spathulenol are three significant terpenoid compounds found in date palm spathe (Taroonah), representing the alcohol, phenol, and alcohol sub-classes, respectively [80]. Linalool exhibits significant anticancer activity through numerous specific pathways that may open avenues for therapeutic applications. This monoterpene can induce apoptosis in various cancer cells by activating intrinsic apoptotic pathways, evidenced by increased expression of pro-apoptotic factors and decreased levels of anti-apoptotic proteins. Linalool may also impact cell cycle regulation, resulting in arrest at critical phases, ultimately inhibiting proliferation in multiple cancer types. Furthermore, its potential as a chemosensitizing agent suggests that linalool can enhance the efficacy of existing chemotherapy treatments, offering a promising approach to improving cancer therapy outcomes [81-83]. Carvacrol may exert its anticancer effects by modulating key signaling pathways and protein expressions involved in tumor progression. It can downregulate matrix metalloproteinase-2 and -9, pivotal in cancer cell invasion and metastasis. Furthermore, carvacrol induces apoptosis through increased pro-apoptotic protein expression, while decreasing levels of anti-apoptotic proteins such as Bcl-2, thereby disrupting the balance of cell survival. Additionally, it may inhibit the phosphoinositide 3-kinase/protein kinase B pathway, and extracellular signal-regulated kinase 1/2 mitogen-activated protein kinase signaling, alongside promoting the phosphorylation of c-Jun N-terminal kinase and p38 MAPK, which collectively influence cancer cell fate and growth [84]. Spathulenol may exert a significant impact on cancer and tumor pathways, particularly in melanoma. It can modulate the MAPK/ERK pathway, potentially leading to decreased cell proliferation and enhanced apoptosis in SK-MEL-28 cells through mechanisms that may involve alterations in key protein expression. Furthermore, spathulenol may disrupt the BRAF signaling pathway, thus potentially reducing cell viability and contributing to the overall inhibition of tumor growth. By promoting intrinsic apoptotic mechanisms, it can enhance the expression of pro-apoptotic proteins, reinforcing its role as a promising candidate in melanoma treatment [85, 86].

The Flowers

Extracts (especially alkaline extract) obtained from the powder of date male flowers show antioxidant properties; Xylose is a monosaccharide that is significantly found in these extracts [87]; Xylitol and xylulose are derivatives of xylose [88]. Xylitol may selectively trigger cancer cell death by activating specific anticancer pathways, potentially inducing apoptosis through the upregulation of CHAC1, which can lead to oxidative stress and promote cell death. This action might involve the induction of endoplasmic reticulum stress, facilitated by ATF4, providing a selective mechanism that spares normal cells while affecting cancerous ones. Furthermore, xylitol may enhance chemotherapeutic efficacy by sensitizing cancer cells to agents like 5-fluorouracil, thereby proposing a supportive role in cancer therapies [89]. Similarly, xylulose is suggested to possess anticancer properties, particularly against colorectal cancer, as it may induce apoptosis by upregulating pro-apoptotic proteins while downregulating anti-apoptotic factors like BCL-2, with a mechanism that can attenuate the MAPK signaling pathway and reduce the phosphorylation of JNK, ERK, and P38, positioning it as a promising agent for targeting tumor growth [90]. The extract

from male date palm flowers exhibited significant antioxidant properties as evidenced by its effectiveness in various assays, including the scavenging of DPPH radicals, enhancement of ferric reducing power, chelation of ferrous ions, and inhibition of β -carotene bleaching, indicating its potential utility in cancer prevention and treatment strategies [91].

The Pollen

Date palm pollen contains bioactive compounds such as caffeic acid, triggering cancer cell apoptosis [92, 93]. Isorhamnetin in this pollen induces antitumor effects through the PI3K/AKT signaling cascade [94, 95]. Hesperidin, a flavonoid in this pollen, can activate caspases and deactivate kinases, compelling cancer cells to undergo apoptosis [96, 97]. Pyrogallol and ellagic acid, both found in date palm pollen, have shown significant promise in cancer treatment. Pyrogallol can reduce tumor size and has a similar effect to the drug doxorubicin on cancer cells, while ellagic acid can inhibit the proliferation and metastasis of tumor cells by regulating PI3K/Akt signaling pathway, JNK (cJun) signaling pathway, mitochondrial pathway, Bcl-2/Bax signaling pathway, TGF- β /Smad3 signaling pathway [98-100]. Date palm pollen contains a complex of fatty acids, with Palmitic acid being the main component [101]. Palmitic acid has been found to increase the expression of caspases and p53 while decreasing the expression of Bcl-2 [102]. As a result, this fatty acid may help prevent the occurrence and spread of cancer cell metastasis [103]. In general, the extract of date palm pollen can reduce the damage caused by radiation therapy and chemotherapy and also play a role in the cancer treatment process [104, 105].

Anticancer Properties of Date Fruits

Date fruit is rich in Lysine and Phenylalanine, essential amino acids [106]. Lysine reduces prostate cancer cell metastasis by reducing acidosis in and around the tumor, enhances Doxorubicin's effect on breast cancer cells, and can increase hydrogen peroxide levels to halt tumor growth [107-109]. Phenylalanine is used as a drug carrier in targeted cancer treatment [110]. A dipeptide derived from this amino acid has been found to inhibit the growth and metastasis of prostate cancer cells by regulating the expression of ANFSF9 and DUSP1 genes [111]. B vitamins are effective anticancer compounds by preventing the disproportionate increase in DNA methylation and breaks in important genes such as P53 and Apc [112]. However, this amount in date fruit varies during ripening and between different cultivars [113]. Selenium is one of the important minerals that have different amounts in different varieties of dates. The significant selenium in date fruits is involved in antioxidation, regulation of the cell cycle, and apoptosis [114, 115]. In the progression of cancer, there is typically a decrease in the levels of superoxide dismutase, glutathione reductase, glutathione peroxidase, and catalase, while the levels of alanine aminotransferase, aspartate aminotransferase, and alkaline phosphatase tend to increase. It has been observed that the aqueous extract of date fruit can reverse both of these processes [116]. Beta D-glucan found in date fruit can be introduced as an antitumor by amplifying phagocytic killing of iC3b-opsonized tumor cells [117, 118]. The bioactive compounds in this fruit aid in the treatment of cancer by inhibiting cancer-causing enzymes like cytochrome p450 and by boosting the activity of anticancer enzymes such as phase II enzymes [119]. Date fruits contain various compounds, including flavonoids, phenols, and carotenoids [120]. Three important flavonoids are Quercetin, Luteolin, and Apigenin [121]. Quercetin activates caspase III, promoting apoptosis and autophagy of cancer cells [122]. Luteolin induces cell cycle arrest

and apoptosis in cancer cell lines by involving DNA damage, regulation of redox, and protein kinases in inhibiting cancer cell proliferation [123]. Apigenin functions by regulating processes associated with cancer suppression, such as the decrease of P13K and MMP and the increase of Bax, p53, and p16 [124]. Date fruits contain quinic acid, rutin, and epicatechin, which are potent anticancer phenolic compounds [125]. Quinic acid reduces the expression of cyclin D1, inhibiting oral cancer cell proliferation [126]. Rutin induces cell cycle arrest in cancer cells by creating changes such as reducing DNA damage and proliferation and increasing caspases and p53; in contrast, epicatechin increases cancer cell sensitivity to radiation and chemotherapy [127, 128]. So far, date fruit extract has been proven to have anticancer properties in multiple independent studies [129-131].

DISCUSSION

Many of the bioactive and anticancer compounds are similar across different organs of the date palm; each organ can also contain unique compounds. The maturation process of the date fruit is characterized by a reduction in its carotenoid levels,

resulting in the mature fruit containing significantly lower concentrations than its seeds. In contrast, compounds such as 2-hydroxycinnamic acid, 4-hydroxycinnamic acid, quercetin, and luteolin are found in greater quantities in the seeds compared to the fruits. Ferulic acid is present in higher concentrations in pollen than in seeds, and seeds contain more ferulic acid than fruits. Catechin levels are more pronounced in the leaves than in the fruits, while the fruits exhibit higher catechin content than the seeds. The concentration of apigenin is greater in pollen than in the fruit, and the fruit also contains more caffeic acid than the seeds [132]. This widespread distribution of bioactives across the entire tree suggests that, similar to the fruit, other organs hold significant potential for valorization in the nutraceutical and pharmaceutical industries [133]. The promising anticancer activities of root endophytes further underscore the potential hidden within the underutilized parts of the date palm [53]. Many *in vivo* studies have been conducted on the effect of extracts taken from different organs of the palm in line with their inhibitory role on cancer-causing agents [134-137]. This indicates that lesser-studied organs are not merely alternative sources but can offer unique therapeutic profiles.

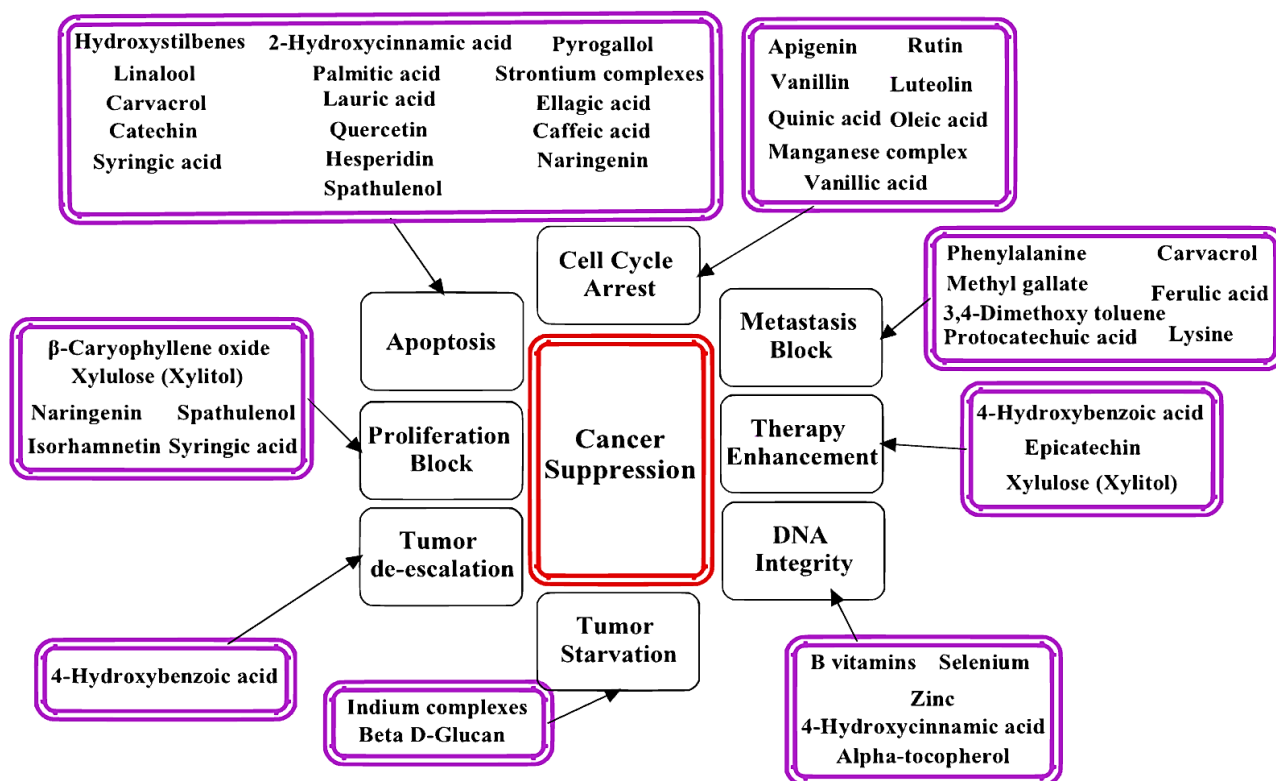


Fig. 2 Pathways of cancer-suppressive action of bioactive compounds of different organs of date palm.

CONCLUSION

It is difficult to compare the amount of bioactive compounds of all the organs in this tree because current studies are more focused on their differences in different cultivars rather than in different organs of the same cultivar. Review suggests that date palms have a high potential in cancer treatment, and their products could be beneficial as a primary or additional treatment. The collective evidence positions the date palm as a comprehensive source of diverse anticancer agents. In Figure 2, the effect of bioactive compounds of different organs of the date palm is reviewed. However, further

research is needed to fully understand the role of all effective substances in date palm organs; A direct comparative analysis of bioactive compounds across all organs from a single cultivar is critically needed. In Figure 3, the potential of six different organs is reviewed. Future research should focus on isolating new compounds from less-studied date palm organs and evaluating their synergistic effects with conventional chemotherapeutic agents. Utilizing these underutilized organs could pave the way for innovative and accessible cancer treatments.

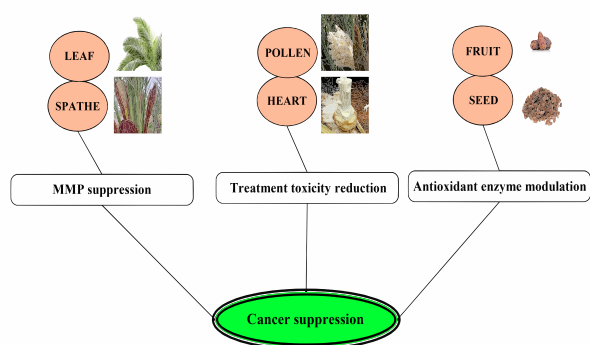


Fig. 3 Potential of six different organs of the date palm in suppressing cancer.

REFERENCES

- Cairns J. The treatment of diseases and the war against cancer. Scientific American. 1985;253(5):51-59.
- Buyel J. Plants as sources of natural and recombinant anti-cancer agents. Biotechnology Advances. 2018;36(2):506-20.
- Lim T. Edible Medicinal and Non-Medicinal Plants (1 ed. Vol. 1): Springer. 2012; 407-418.
- Nixon R.W. Growing dates in the United States. United States Department of Agriculture: Agricultural Research Service. 1959; 3-4.
- Krueger R.R. Date palm (*Phoenix dactylifera* L.) biology and utilization The Date Palm Genome. Phylogeny, Biodiversity and Mapping. 2021;1:3-28.
- Keskin B., Uzundumlu A.S., Karabacak T. Production Estimates of Leading Countries in Date Production for the Period 2023–2028. Applied Fruit Science. 2025;67(2):83.
- Saboori S., Noormohammadi Z., Sheidai M., Marashi S. Date palm (*Phoenix dactylifera* L.) cultivar relationships based on chloroplast genotyping. Iranian Journal of Science and Technology, Transactions A: Science. 2021;45(3):833-40.
- Hartwell J.L. Plants used against cancer: A Survey. I-XI. Lloydia. 1967-1971; 30.
- Nazri M., Yusoff A.M., Nazri N.F.Z., Khalaf N.A.R., Rahman A.A. The descriptions of date palms and an ethnomedicinal importance of dates mentioned in the quran. Mediterranean J of Social Sciences. 2016;7(2):405-17.
- Chao C.T., Krueger R.R. The date palm (*Phoenix dactylifera* L.): overview of biology, uses, and cultivation. HortScience. 2007;42(5):1077-82.
- Sadeghi Z., Kuhestani K. Ethnobotany of date palm (*Phoenix dactylifera*) in Baluch tribe of Saravan region, Baluchistan, Iran. 2014.
- Hamini F., Yousfi M., Soumaya H. Phytochemical analysis, antioxidant properties and phenolic acids determination of date palm (*Phoenix dactylifera* L.) leaves and roots extracts. Journal of the Chilean Chemical Society. 2025;70(2):6330-35.
- Karimi M., Asbagh F.A., Safavi M., Yekaninejad M.S., Rahimi R., Pourmand G., Mirzaei M., Farshbaf-Khalili A. The Effect of Date Palm (*Phoenix dactylifera* L.) Pollen on the Serum Levels of Testosterone, Luteinizing, and Follicle Stimulating Hormones in Men. Current Traditional Medicine. 2024;10(3):101-06.
- Nwachukwu S.C., Edo G.I., Samuel P.O., Jikah A.N., Oloni G.O., Ezekiel G.O., Agbo J.J. The botanical details, pharmacological activities and industrial applications of date seed (*Phoenix dactylifera* L.). Phytochemistry Reviews. 2025;24(1):927-51.
- Elkot W.F., Alsulami T., Malek G., Abo-Srea M.M., Abdallah S.E., Al-Farga A., Elmahdy A. Effect of using heart of date palm as a new source of protein, carbohydrates, and bioactive compounds on the characterization of low-fat camel milk ice cream. International Journal of Biological Macromolecules. 2025;289:138869.
- Asadi K., Safaeian R., Babaei S., Niakousari M. A Comparative Study of Phenolic, Flavonoid and Antioxidant Compounds for Spathe and Male Flower of *Phoenix dactylifera* L.: Extraction Method and Solvent Effects. Applied Fruit Science. 2025;67(1):32.
- Zarie A.A., Hassan A.B., Alshammari G.M., Yahya M.A., Osman M.A. Date industry by-product: date seeds (*Phoenix dactylifera* L.) as potential natural sources of bioactive and antioxidant compounds. Applied Sciences. 2023;13(21):11922.
- Benincasa C., Nicoletti R., Harkat H., Bousba R., Atrouz K., Gültekin-Özgüven M., Altuntaş Ü., Demircan E., Zahran H.A., Özçelik B. Composizione biochimica e proprietà antiossidanti dell'olio di semi di dattero (*Phoenix dactylifera* L.). Rivista Italiana delle Sostanze Grasse. 2023;100(3):157-64.
- Bezerra D.P., Soares A.K.N., de Sousa D.P. Overview of the role of vanillin on redox status and cancer development. Oxidative Medicine and Cellular Longevity. 2016;2016(1):9734816.
- Deguchi H., Fujii T., Nakagawa S., Koga T., Shirouzu K. Analysis of cell growth inhibitory effects of catechin through MAPK in human breast cancer cell line T47D. International Journal of Oncology. 2002;21(6):1301-05.
- Al-Hazzani A.A., Alshatwi A.A. Catechin hydrate inhibits proliferation and mediates apoptosis of SiHa human cervical cancer cells. Food and Chemical Toxicology. 2011;49(12):3281-86.
- Przystupski D., Michel O., Rossowska J., Kwiatkowski S., Saczko J., Kulbacka J. The modulatory effect of green tea catechin on drug resistance in human ovarian cancer cells. Medicinal Chemistry Research. 2019;28:657-67.
- Alkhalaf M.I., Churchill G.C., Mirghani M.E. Chemical composition and antioxidant/antibacterial depictions of Zahidi date palm (*Phoenix dactylifera*) kernel oil. Journal of King Saud University-Science. 2023;35(7):102817.
- Mercado Y.G., Díaz J.M., Hernández D.O., Gonzalez F.L., Zaldivar E.R., Sapiens M.H., Pinedo U.G., Estrada R., Carballo M.M., Aguirre A.C. Ortho-coumaric acid derivatives with therapeutic potential in a three-dimensional culture of the immortalised U-138 MG glioblastoma multiforme cell line. Neurology Perspectives. 2022;2(1):S19-S30.
- Gong J., Zhou S., Yang S. Vanillic Acid Suppresses HIF-1 α Expression via Inhibition of mTOR/p70S6K/4E-BP1 and Raf/MEK/ERK Pathways in Human Colon Cancer HCT116 Cells. International Journal of Molecular Sciences. 2019;20(3).
- Kaur J., Gulati M., Gowthamarajan K., Vishwas S., Chellappan D.K., Gupta G., Dua K., Pandey N.K., Kumar B., Singh S.K. Combination therapy of vanillic acid and oxaliplatin co-loaded in polysaccharide based functionalized polymeric micelles could offer effective treatment for colon cancer: A hypothesis. Medical Hypotheses. 2021;156:110679.
- Park J., Cho S.Y., Kang J., Park W.Y., Lee S., Jung Y., Kang M.-W., Kwak H.J., Um J.-Y. Vanillic acid improves comorbidity of cancer and obesity through stat3 regulation in high-fat-diet-induced obese and b16b16 melanoma-injected mice. Biomolecules. 2020;10(8):1098.
- Seidel C., Schneckeburger M., Mazumder A., Teiten M.-H., Kirsch G., Dicato M., Diederich M. 4-Hydroxybenzoic acid derivatives as HDAC6-specific inhibitors modulating microtubular structure and HSP90 α chaperone activity against prostate cancer. Biochemical Pharmacology. 2016;99(1):31-52.
- Wang X.-N., Wang K.-Y., Zhang X.-S., Yang C., Li X.-Y. 4-Hydroxybenzoic acid (4-HBA) enhances the sensitivity of human breast cancer cells to adriamycin as a specific HDAC6 inhibitor by promoting HIPK2/p53 pathway. Biochemical and Biophysical Research Communications. 2018;504(4):812-19.
- Faridi N., Ghahghaei A. Hsp90 structure and function in cancer. Gene, Cell and Tissue. 2018;5(3).
- Niero E.L.d.O., Machado-Santelli G.M. Cinnamic acid induces apoptotic cell death and cytoskeleton disruption in human melanoma cells. Journal of Experimental & Clinical Cancer Research. 2013;32:1-14.
- Calori I.R., Piva H.L., Tedesco A.C. Targeted cancer therapy using alpha-cyano-4-hydroxycinnamic acid as a novel vector molecule: A proof-of-concept study. Journal of Drug Delivery Science and Technology. 2020;57:101633.
- Laghouiter O.K., Benalia M., Gourine N., Djeridane A., Bombarda I., Yousfi M. Chemical characterization and in vitro antioxidant capacity of nine Algerian date palm cultivars (*Phoenix dactylifera* L.) seed oil. Mediterranean Journal of Nutrition and Metabolism. 2018;11(2):103-17.
- Das Gupta S., Suh N. Tocopherols in cancer: An update. Molecular Nutrition & Food Research. 2016;60(6):1354-63.

35. Besbes S., Blecker C., Deroanne C., Drira N.-E., Attia H. Date seeds: chemical composition and characteristic profiles of the lipid fraction. Food Chemistry. 2004;84(4):577-84.
36. Karacor K., Cam M. Effects of oleic acid. Medical Science and Discovery. 2015;2(1):125-32.
37. Deng B., Kong W., Suo H., Shen X., Newton M.A., Burkett W.C., Zhao Z., John C., Sun W., Zhang X. Oleic acid exhibits anti-proliferative and anti-invasive activities via the PTEN/AKT/mTOR pathway in endometrial cancer. Cancers. 2023;15(22):5407.
38. Mustafa A., Indiran M.A., Ramalingam K., Perumal E., Shanmugham R., Karobari M.I. Anticancer potential of thiocolchicoside and lauric acid loaded chitosan nanogel against oral cancer cell lines: a comprehensive study. Scientific Reports. 2024;14(1):9270.
39. Hilary S., Kizhakkayil J., Souka U., Al-Meqbaali F., Ibrahim W., Platat C. In-vitro investigation of polyphenol-rich date (*Phoenix dactylifera* L.) seed extract bioactivity. Frontiers in Nutrition. 2021;8(1):667514.
40. Habib H.M., El-Fakharany E.M., Souka U.D., Elsebae F.M., El-Ziney M.G., Ibrahim W.H. Polyphenol-rich date palm fruit seed (*Phoenix dactylifera* L.) extract inhibits labile iron, enzyme, and cancer cell activities, and DNA and protein damage. Nutrients. 2022;14(17):3536.
41. Habib H.M., Ibrahim W.H. Effect of date seeds on oxidative damage and antioxidant status in vivo. Journal of the Science of Food and Agriculture. 2011;91(9):1674-79.
42. Rezaei M., Khodaei F., Hooshmand N. Date seed extract diminished apoptosis event in human colorectal carcinoma cell line. MOJ Toxicol. 2015;1(4):00017.
43. Khan M.A., Singh R., Siddiqui S., Ahmad I., Ahmad R., Upadhyay S., Barkat M.A., Ali A.M.A., Zia Q., Srivastava A. Anticancer potential of *Phoenix dactylifera* L. seed extract in human cancer cells and pro-apoptotic effects mediated through caspase-3 dependent pathway in human breast cancer MDA-MB-231 cells: an in vitro and in silico investigation. BMC Complementary Medicine and Therapies. 2022;22(1):68.
44. Alwattar M., Shalan N., Alaraj M. Anticancer effects of date seed oil-loaded niosomes and gemcitabine combination in MCF-7 human breast cancer cells. Research Journal of Pharmacy and Technology. 2023;16(9):4179-87.
45. Benabbes R., Ouahhoud S., Moueqqit M., Addi M., Hano C., Delporte C., Nacoulma A.P., Megalizzi V. The Major Stilbene Compound Accumulated in the Roots of a Resistant Variety of *Phoenix dactylifera* L. Activates Proteasome for a Path in Anti-Aging Strategy. Cells. 2022;12(1):71.
46. Sirerol J.A., Rodríguez M.L., Mena S., Asensi M.A., Estrela J.M., Ortega A.L. Role of natural stilbenes in the prevention of cancer. Oxidative Medicine and Cellular Longevity. 2016;2016(1):3128951.
47. Al-Abdoulhadi I., Dinar H., Ebert G., Büttner C. Influence of salinity levels on nutrient content in leaf, stem and root of major date palm (*Phoenix dactylifera* L.) Cultivars. 2012;2(8):341-46.
48. Jacobs M.M., Pienta R.J. Relationships between potassium and cancer Vitamins and Minerals in the Prevention and Treatment of Cancer (pp. 227-45): CRC Press. 2018.
49. Cheemanapalli Srinivasulu C.S., Mopuri Ramgopal M.R., Golla Ramanjaneyulu G.R., Anuradha C., Kumar C. Syringic acid (SA)-a review of its occurrence, biosynthesis, pharmacological and industrial importance. 2018.
50. Pei J., Velu P., Zareian M., Feng Z., Vijayalakshmi A. Effects of syringic acid on apoptosis, inflammation, and AKT/mTOR signaling pathway in gastric cancer cells. Frontiers in Nutrition. 2021;8:788929.
51. Gao J., Yu H., Guo W., Kong Y., Gu L., Li Q., Yang S., Zhang Y., Wang Y. The anticancer effects of ferulic acid is associated with induction of cell cycle arrest and autophagy in cervical cancer cells. Cancer Cell International. 2018;18:1-9.
52. Bao X., Li W., Jia R., Meng D., Zhang H., Xia L. Molecular mechanism of ferulic acid and its derivatives in tumor progression. Pharmacological Reports. 2023;75(4):891-906.
53. Ben Mefteh F., Daoud A., Chenari Bouket A., Thissera B., Kadri Y., Cherif-Silini H., Eshelli M., Alenezi F.N., Vallat A., Oszako T. Date palm trees root-derived endophytes as fungal cell factories for diverse bioactive metabolites. International Journal of Molecular Sciences. 2018;19(7):1986.
54. Oves M., Aslam M., Rauf M.A., Qayyum S., Qari H.A., Khan M.S., Alam M.Z., Tabrez S., Pugazhendhi A., Ismail I.M. Antimicrobial and anticancer activities of silver nanoparticles synthesized from the root hair extract of *Phoenix dactylifera*. Materials Science and Engineering: C. 2018;89:429-43.
55. Trabzuni D.M., Ahmed S.E.B., Abu-Tarboush H.M. Chemical composition, minerals and antioxidants of the heart of date palm from three Saudi cultivars. Food and Nutrition Sciences. 2014;5(14):1379.
56. Ajiboye T.O., Amao I.O., Adeyemi W.J., Babalola S.O., Akinsuyi O.S., Ogunrombi M.O., Ogunlaja A.S., Mhlanga S.D. Overview of medical and biological applications of indium (iii) complexes. Chemistry Africa. 2024;7(4):1729-48.
57. Khaksar S., Panjehpour A., Ghadermazi E., Motieyan E., Aliabadi A., Rostamnia S., Marabello D., Abdolmaleki S. Study on crystallographic structure and antiproliferative effect of mixed-ligand strontium (II) complex and N, N'-bis (2-hydroxy-5-methylphenyl) pyridine-2, 6-dicarboxamide ligand. Journal of Molecular Structure. 2023;1274:134432.
58. Mostafa M.Y. " Evaluation of Antioxidant, Anti-Inflammatory, and Anticancer Activities of Palm Heart (*Phoenix dactylifera* L.) In Vitro". Journal of Food & Dairy Sciences. 2024;15(9).
59. Huang S., Wang L.-L., Xue N.-N., Li C., Guo H.-H., Ren T.-K., Zhan Y., Li W.-B., Zhang J., Chen X.-G. Chlorogenic acid effectively treats cancers through induction of cancer cell differentiation. Theranostics. 2019;9(23):6745.
60. Cortez N., Villegas C., Burgos V., Ortiz L., Cabrera-Pardo J.R., Paz C. Therapeutic Potential of Chlorogenic Acid in Chemoresistance and Chemoprotection in Cancer Treatment. International Journal of Molecular Sciences. 2024;25(10):5189.
61. Santana-Gálvez J., Villela Castrejón J., Serna-Saldívar S.O., Jacobo-Velázquez D.A. Anticancer potential of dihydrocaffeic acid: a chlorogenic acid metabolite. CyTA-Journal of Food. 2020;18(1):245-48.
62. Choi J., Lee D.-H., Jang H., Park S.-Y., Seol J.-W. Naringenin exerts anticancer effects by inducing tumor cell death and inhibiting angiogenesis in malignant melanoma. International Journal of Medical Sciences. 2020;17(18):3049.
63. Stabrauskienė J., Kopustinskiene D.M., Lazauskas R., Bernatoniene J. Naringin and naringenin: Their mechanisms of action and the potential anticancer activities. Biomedicines. 2022;10(7):1686.
64. Sahyon H.A., Al-Harbi S.A. Chemoprotective role of an extract of the heart of the *Phoenix dactylifera* tree on adriamycin-induced cardiotoxicity and nephrotoxicity by regulating apoptosis, oxidative stress and PD-1 suppression. Food and Chemical Toxicology. 2020;135:111045.
65. Perveen K., Bokhari N.A., Dina A. Analysis of essential elements of three different varieties of Saudi Arabian date palm (*Phoenix dactylifera*). Asian Journal of Chemistry. 2013;25(9):5092.
66. Dhawan D., Chadha V.D. Zinc: a promising agent in dietary chemoprevention of cancer. Indian Journal of Medical Research. 2010;132(6):676-82.
67. Irfandi R., Raya I. Potential anticancer activity of Mn (II) complexes containing arginine dithiocarbamate ligand on MCF-7 breast cancer cell lines. Annals of Medicine and Surgery. 2020;60:396-402.
68. Laouini S., Segni L., Ouahrani M., Gherraf N., Mokni S. Phytochemical analysis, antioxidant and antimicrobial activities of leaves extract of date palm grown in Algeria. Journal of Fundamental and Applied Sciences. 2012;4(2):142-54.
69. John J.A., Shahidi F. Phenolic content, antioxidant and anti-inflammatory activities of seeds and leaves of date palm (*Phoenix dactylifera* L.). Journal of Food Bioactives. 2019;5:120-30-20-30.
70. Liang H., Chen Z., Yang R., Huang Q., Chen H., Chen W., Zou L., Wei P., Wei S., Yang Y. Methyl Gallate Suppresses the Migration, Invasion, and Epithelial-Mesenchymal Transition of Hepatocellular Carcinoma Cells via the AMPK/NF-κB Signaling Pathway in vitro and in vivo. Frontiers in Pharmacology. 2022;13:894285.
71. Lin H.H., Chen J.H., Chou F.P., Wang C.J. Protocatechuic acid inhibits cancer cell metastasis involving the down-regulation of Ras/Akt/NF-κB pathway and MMP-2 production by targeting RhoB activation. British Journal of Pharmacology. 2011;162(1):237-54.
72. Punvittayagul C., Luangsaphabool T., Wongpoomchai R. Protocatechuic acid as a potent anticarcinogenic compound in purple rice bran against diethylnitrosamine-initiated rat hepatocarcinogenesis. Scientific Reports. 2022;12(1):10548.

73. Chakroun M., Morjen M., Mabrouk H.B., Mejdoub H., Srairi-Abid N., Marrakchi N., Jebali J., Khemakhem B. Anticancer Properties of Different Varieties of Date Palm (*Phoenix dactylifera* L.) Leaf Extracts in Human Tumor Cells: a Comparative Study. *Plant Foods for Human Nutrition*. 2024;79(2):518–25.
74. Mard S.A., Jalalvand K., Jafarinejad M., Balochi H., Naseri M.K.G. Evaluation of the antidiabetic and antilipemic activities of the hydroalcoholic extract of *Phoenix dactylifera* palm leaves and its fractions in alloxan-induced diabetic rats. *The Malaysian Journal of Medical Sciences: MJMS*. 2010;17(4):4.
75. Atghaei M., Sefidkon F., Darini A., Sadeghzadeh Hemayati S., Abdossi V. Essential oil content and composition of the spathe in some date palm (*Phoenix dactylifera* L.) varieties in Iran. *Journal of Essential Oil Bearing Plants*. 2020;23(2):292–300.
76. Al-Taher A.Y. Anticonvulsant effects of 3, 4-Dimethoxy toluene, the major constituent of *Phoenix dactylifera* L Spathe in mice. *Scientific Journal of King Faisal University (Basic and Applied Sciences)*. 2008;9(2):115–23.
77. Ngo D.-H., Vo T.S. An updated review on pharmaceutical properties of gamma-aminobutyric acid. *Molecules*. 2019;24(15):2678.
78. Farbodniaye Jahromi M.A., Moien M.R., Mollaei M. Volatile Constituents and Antioxidant Activity of Spathes from Five Uncommon Varieties of *Phoenix dactylifera* L. *Trends in Pharmaceutical Sciences*. 2018;4(4):225–34.
79. Fidyat K., Fiedorowicz A., Strzdała L., Szumny A. β -caryophyllene and β -caryophyllene oxide—natural compounds of anticancer and analgesic properties. *Cancer Medicine*. 2016;5(10):3007–17.
80. Hamed A., Mohagheghzadeh A., Rivaz S. Preliminary pharmacognostic evaluation and volatile constituent analysis of spathe of *Phoenix dactylifera* L. (Tarooneh). *Pharmacognosy Journal*. 2013;5(2):83–86.
81. Pereira I., Severino P., Santos A.C., Silva A.M., Souto E.B. Linalool bioactive properties and potential applicability in drug delivery systems. *Colloids and Surfaces B: Biointerfaces*. 2018;171:566–78.
82. Zhao Y., Cheng X., Wang G., Liao Y., Qing C. Linalool inhibits 22Rv1 prostate cancer cell proliferation and induces apoptosis. *Oncology Letters*. 2020;20(6):1–1.
83. Elbe H., Ozturk F., Yigitturk G., Baygar T., Cavusoglu T. Anticancer activity of linalool: Comparative investigation of ultrastructural changes and apoptosis in breast cancer cells. *Ultrastructural Pathology*. 2022;46(4):348–58.
84. Imran M., Aslam M., Alsagaby S.A., Saeed F., Ahmad I., Afzaal M., Arshad M.U., Abdelgawad M.A., El-Ghorab A.H., Khamis A. Therapeutic application of carvacrol: A comprehensive review. *Food Science & Nutrition*. 2022;10(11):3544–61.
85. Beserra Santos L.K., Alves Veras M.D., Gadelha Marques K.K., De Moraes Alves M.M., Mendes A.N., De Amorim Carvalho F.A., Sobral M.V., Chaves M.H., Ramos Goncalves J.C. Assessment of In Vitro Antimelanoma Potential of Ephedranthus pisocarpus REFr. *Anticancer Research*. 2020;40(9):5015–24.
86. Bomfim L.M., Menezes L.R., Rodrigues A.C.B., Dias R.B., Gurgel Rocha C.A., Soares M.B., Neto A.F., Nascimento M.P., Campos A.F., Silva L.C.e. Antitumour activity of the microencapsulation of Annona vepretorum essential oil. *Basic & Clinical Pharmacology & Toxicology*. 2016;118(3):208–13.
87. Karra S., Sebii H., Yaich H., Bouaziz M.A., Blecker C., Danthine S., Attia H., Besbes S. Effect of extraction methods on the physicochemical, structural, functional, and antioxidant properties of the dietary fiber concentrates from male date palm flowers. *Journal of Food Biochemistry*. 2020;44(6):e13202.
88. Meng Q., Zhang T., Jiang B., Mu W., Miao M. Advances in applications, metabolism, and biotechnological production of L-xylulose. *Applied Microbiology and Biotechnology*. 2016;100:535–40.
89. Tomonobu N., Komalasari N.L.G.Y., Sumardika I.W., Jiang F., Chen Y., Yamamoto K.-i., Kinoshita R., Murata H., Inoue Y., Sakaguchi M. Xylitol acts as an anticancer monosaccharide to induce selective cancer death via regulation of the glutathione level. *Chemico-biological interactions*. 2020;324:109085.
90. Hu Q., Zheng Q., Du X., Yang Z., Tian Q., Liang L., Zhao X., Bai H., Liu Y., Zhao M. Intestinal metabolite xylulose inhibits colorectal cancer by inducing apoptosis through the MAPK signalling pathway. *Toxicology and Applied Pharmacology*. 2024;487:116960.
91. Karra S., Sebii H., Jarak M., Bouaziz M.A., Attia H., Blecker C., Besbes S. Male date palm flowers: Valuable nutritional food ingredients and alternative antioxidant source and antimicrobial agent. *South African Journal of Botany*. 2020;131:181–87.
92. Daoud A., Malika D., Bakari S., Hfaiedh N., Mnafigui K., Kadri A., Gharsallah N. Assessment of polyphenol composition, antioxidant and antimicrobial properties of various extracts of Date Palm Pollen (DPP) from two Tunisian cultivars. *Arabian Journal of Chemistry*. 2019;12(8):3075–86.
93. Alam M., Ahmed S., Elasbali A.M., Adnan M., Alam S., Hassan M.I., Pasupuleti V.R. Therapeutic implications of caffeic acid in cancer and neurological diseases. *Frontiers in Oncology*. 2022;12:860508.
94. Al-Samarrai R.R., Al-Samarrai A.-M.H., Al-Salihi F.G. Identification of flavonoids in Iraqi date palm pollen by HPLC. *Orient. J. Chem*. 2017;33(2):985–88.
95. Zhai T., Zhang X., Hei Z., Jin L., Han C., Ko A.T., Yu X., Wang J. Isorhamnetin inhibits human gallbladder cancer cell proliferation and metastasis via PI3K/AKT signaling pathway inactivation. *Frontiers in Pharmacology*. 2021;12:628621.
96. Abdel-Shaheed M.M., Abdalla E.S., Khalil A.F., El-Hadidy E.M. Effect of Egyptian date palm pollen (*Phoenix dactylifera* L.) and its hydroethanolic extracts on serum glucose and lipid profiles in induced diabetic rats. *Food and Nutrition Sciences*. 2021;12(2):147–61.
97. Aggarwal V., Tuli H.S., Thakral F., Singhal P., Aggarwal D., Srivastava S., Pandey A., Sak K., Varol M., Khan M.A. Molecular mechanisms of action of hesperidin in cancer: Recent trends and advancements. *Experimental Biology and Medicine*. 2020;245(5):486–97.
98. Abdallah W.E., Awad H.M., AbdelMohsen M.M. Phytochemical Composition, Antioxidant and Antitumor Activities of some Date Palm Pollen Extracts. *Egyptian Journal of Chemistry*. 2023;66(5):425–34.
99. Revathi S., Hakkim F.L., Kumar N.R., Bakshi H.A., Sangilimuthu A.Y., Tambuwala M.M., Changez M., Nasef M.M., Krishnan M., Kayalvizhi N. In vivo anti cancer potential of pyrogallol in murine model of colon cancer. *Asian Pacific Journal of Cancer Prevention: APJCP*. 2019;20(9):2645.
100. Lu G., Wang X., Cheng M., Wang S., Ma K. The multifaceted mechanisms of ellagic acid in the treatment of tumors: State-of-the-art. *Biomedicine & Pharmacotherapy*. 2023;165:115132.
101. Hassan H.M. Chemical composition and nutritional value of palm pollen grains. *Global J Biotechnol Biochem*. 2011;6(1):1–7.
102. Zafaryab M., Fakhri K.U., Khan M.A., Hajela K., Rizvi M.M.A. In vitro assessment of cytotoxic and apoptotic potential of palmitic acid for breast cancer treatment. *International Journal Life Science Research*. 2019;7(1):166–74.
103. Yu X., Peng W., Wang Y., Xu W., Chen W., Huang L., Xu H., He X., Wang S., Sun Q. Palmitic acid inhibits the growth and metastasis of gastric cancer by blocking the STAT3 signaling pathway. *Cancers*. 2023;15(2):388.
104. Elkerm Y., Tawashi R. Date palm pollen as a preventative intervention in radiation-and chemotherapy-induced oral mucositis: a pilot study. *Integrative Cancer Therapies*. 2014;13(6):468–72.
105. Elblehi S.S., El-Sayed Y.S., Soliman M.M., Shukry M. Date palm pollen extract avert doxorubicin-induced cardiomyopathy fibrosis and associated oxidative/nitrosative stress, inflammatory cascade, and apoptosis-targeting Bax/Bcl-2 and Caspase-3 signaling pathways. *Animals*. 2021;11(3):886.
106. Shaba E.Y., Ndamitso M.M., Tsado J.M., Etsunyakpa M.B., Tsado A.N., Muhammed S.S. Nutritional and anti-nutritional composition of date palm (*Phoenix dactylifera* L.) fruits sold in major markets of Minna Niger State, Nigeria. 2015;9(8):167–74.
107. Ibrahim-Hashim A., Wojtkowiak J.W., Ribeiro M.d.L.C., Estrella V., Bailey K.M., Cornell H.H., Gatenby R.A., Gillies R.J. Free base lysine increases survival and reduces metastasis in prostate cancer model. *Journal of Cancer Science & Therapy*. 2011;1(4):1–4.
108. Jahani M., Yarani R., Rezazadeh D., Tahmasebi H., Hoseinkhani Z., Kiani S., Mansouri K. L-lysine Increases the Anticancer Effect of Doxorubicin in Breast Cancer by Inducing ROS-Dependent Autophagy. *Current Cancer Drug Targets*. 2024;1(24):1–13.
109. Lukasheva E.V., Babayeva G., Karshieva S.S., Zhdanov D.D., Pokrovsky V.S. L-lysine α -oxidase: enzyme with anticancer properties. *Pharmaceuticals*. 2021;14(11):1070.
110. Yin Y., Sheng L., Zhang J., Zhang L., Liu J., Wen X., Liu Y., Si Y., Cheng K. Synthesis and in vitro/in vivo anticancer evaluation of pentacyclic

- triterpenoid derivatives linked with l-phenylalanine or l-proline. *Bioorganic Chemistry*. 2022;126(1):105865.
111. Li L., Yang M., Yu J., Cheng S., Ahmad M., Wu C., Wan X., Xu B., Ben-David Y., Luo H. A novel L-phenylalanine dipeptide inhibits the growth and metastasis of prostate cancer cells via targeting DUSP1 and TNFSF9. *International Journal of Molecular Sciences*. 2022;23(18):10916.
 112. Plazar N., Jurdana M. Hyperhomocysteinemia and the role of B vitamins in cancer. *Radiology and Oncology*. 2010;44(2):79-85.
 113. Aljaloud S., Collieran H.L., Ibrahim S.A. Nutritional value of date fruits and potential use in nutritional bars for athletes. *Food and Nutrition Sciences*. 2020;11(6):463-80.
 114. Al-Showiman S.S., Al-Tamrah S.A., BaOsman A.A. Determination of selenium content in dates of some cultivars grown in Saudi Arabia. *International Journal of Food Sciences and Utrition*. 1994;45(1):29-33.
 115. Rua R.M., Nogales F., Carreras O., Ojeda M.L. Selenium, selenoproteins and cancer of the thyroid. *Journal of Trace Elements in Medicine and Biology*. 2023;76:127115.
 116. Khan F., Khan T.J., Kalamegam G., Pushparaj P.N., Chaudhary A., Abuzenadah A., Kumosani T., Barbour E., Al-Qahtani M. Anti-cancer effects of Ajwa dates (*Phoenix dactylifera* L.) in diethylnitrosamine induced hepatocellular carcinoma in Wistar rats. *BMC Complementary and Alternative Medicine*. 2017;17:1-10.
 117. Ishurd O., Zgheel F., Kermagi A., Flefla M., Elmabruk M. Antitumor activity of β -d-glucan from Libyan dates. *Journal of Medicinal Food*. 2004;7(2):252-55.
 118. Weitberg A.B. A phase I/II trial of beta-(1, 3)/(1, 6) D-glucan in the treatment of patients with advanced malignancies receiving chemotherapy. *Journal of Experimental & Clinical Cancer Research*. 2008;27:1-4.
 119. Diab K., Aboul-Ela E. In vivo comparative studies on antigenotoxicity of date palm (*Phoenix dactylifera* L.) pits extract against DNA damage induced by N-Nitroso-N-methylurea in mice. *Toxicology International*. 2012;19(3):279.
 120. Yasin B.R., El-Fawal H.A., Mousa S.A. Date (*Phoenix dactylifera*) polyphenolics and other bioactive compounds: A traditional islamic remedy's potential in prevention of cell damage, cancer therapeutics and beyond. *International Journal of Molecular Sciences*. 2015;16(12):30075-90.
 121. Al-Shwyeh H.A. Date palm (*Phoenix dactylifera* L.) fruit as potential antioxidant and antimicrobial agents. *Journal of Pharmacy and Bioallied Sciences*. 2019;11(1):1-11.
 122. Reyes-Farias M., Carrasco-Pozo C. The anti-cancer effect of quercetin: molecular implications in cancer metabolism. *International Journal of Molecular Sciences*. 2019;20(13):3177.
 123. Çetinkaya M., Baran Y. Therapeutic potential of luteolin on cancer. *Vaccines*. 2023;11(3):554.
 124. Rahmani A.H., Alsahli M.A., Almatroudi A., Almogbel M.A., Khan A.A., Anwar S., Almatroodi S.A. The potential role of apigenin in cancer prevention and treatment. *Molecules*. 2022;27(18):6051.
 125. Bettaieb I., Kilani A., Ben Othman K., Benabderrahim M.A., Elfalleh W. Phenolic profile, sugar composition, and antioxidant capacities of some common date palm (*Phoenix dactylifera* L.) cultivars as a potential nutraceutical and functional food ingredients. *Journal of Food Quality*. 2023;2023(1):2474900.
 126. Shay J., Elbaz H.A., Lee I., Zielske S.P., Malek M.H., Hüttemann M. Molecular mechanisms and therapeutic effects of (–)-epicatechin and other polyphenols in cancer, inflammation, diabetes, and neurodegeneration. *Oxidative Medicine and Cellular Longevity*. 2015;2015(1):181260.
 127. Nouri Z., Fakhri S., Nouri K., Wallace C.E., Farzaei M.H., Bishayee A. Targeting multiple signaling pathways in cancer: The rutin therapeutic approach. *Cancers*. 2020;12(8):2276.
 128. Abdulkhaleq L.A., Assi M.A., Noor M.H.M., Abdullah R., Saad M.Z., Taufiq-Yap Y.H. Therapeutic uses of epicatechin in diabetes and cancer. *Veterinary world*. 2017;10(8):869.
 129. Rozila I., Abdul Manap N., Ghazali L.N., Kamal N., Abdul Hakeem W., Mohd Manzor N., Chowdhury S.R., Abdul Rahman S. The antioxidant properties and anticancer effect of Medjool dates (*Phoenix dactylifera* L.) on human breast adenocarcinoma (MCF-7) cells: in vitro study. Paper presented at the Front. Pharmacol. Conference Abstract: International Conference on Drug Discovery and Translational Medicine, Putrajaya, Malaysia, 2018.
 130. Shabbaz K., Asif J.A., Liszen T., Nurul A.A., Alam M.K. Cytotoxic and antioxidant effects of *Phoenix dactylifera* L. (Ajwa date extract) on oral squamous cell carcinoma cell line. *BioMed Research International*. 2022;2022(1):5792830.
 131. Elhemeidy R.M.M., Lyrwati D., Widjanto E. Date Fruit Extract (*Phoenix dactylifera*, Ajwa) Modulates NK Cells and TNF-Alpha in DMBA-Induced Mammary Cancer Sprague-Dawley Rats. *Journal of Tropical Life Science*. 2018;8(3):227-35.
 132. Echegaray N., Gullón B., Pateiro M., Amarowicz R., Misihairabgwi J.M., Lorenzo J.M. Date fruit and its by-products as promising source of bioactive components: A review. *Food Reviews International*. 2023;39(3):1411-32.
 133. Ghnimi S., Umer S., Karim A., Kamal-Eldin A. Date fruit (*Phoenix dactylifera* L.): An underutilized food seeking industrial valorization. *NFS Journal*. 2017;6:1-10.
 134. Khalil H.E., Alqahtani N.K., Darrag H.M., Ibrahim H.-I.M., Emeka P.M., Badger-Emeka L.I., Matsunami K., Shehata T.M., Elsewedy H.S. Date palm extract (*Phoenix dactylifera*) PEGylated nanoemulsion: Development, optimization and cytotoxicity evaluation. *Plants*. 2021;10(4):735.
 135. Al-Sayyed H.F., Takruri H.R., Shomaf M.S. The effect of date palm fruit (*Phoenix dactylifera* L.) on 7, 12-dimethylbenz (α) anthracene (DMBA)-Induced Mammary Cancer in Rats. 2014;4(1):11.
 136. Ghanem K.Z., Ramadan M.M., Ghanem H.Z., Fadel M. Improving the production of unsaturated fatty acid esters and flavonoids from date palm pollen and their effects as anti-breast-cancer and antiviral agents: an in-vitro: study. *Journal of The Arab Society for Medical Research*. 2015;10(2):47-55.
 137. Al-Bagoury A., Awad V., Attia L. Anticancer activity of *Phoenix dactylifera* seeds extract: In vitro and in vivo studies. *Biological and Biomedical Journal*. 2023;1(1):17-22.